Comparison of the Neuroprotective Effects of Adrenoceptor Drugs in Retinal Cell Culture and Intact Retina

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PURPOSE. The efficacy of β1-adrenoceptor (AR)-selective (betaxolol and metoprolol) and nonselective (timolol) antagonists and the α2-AR agonist UK14,304 as retinal neuroprotectants was compared and contrasted in an in vitro glutamate excitotoxicity model. The ability of UK14,304, brimonidine, and betaxolol to alter glutamate-receptor-induced changes in intracellular calcium ([Ca2+]i) was also determined in isolated retinal neurons and retinal ganglion cells (RGCs) in an intact retina preparation.

METHODS. Neuronal survival was measured in mixed retinal cell cultures treated for 24 hours with media containing 100 μM glutamate, with or without the addition of each of the drugs (1–1000 μM). Effects of glutamate on C6 glioma cell line. Glutamate-induced changes in [Ca2+]i, with and without UK14,304, and its analogue brimonidine were assessed by calcium-imaging techniques in retinal neurons in culture. The effect of betaxolol on [Ca2+]i was investigated in RGCs in intact rabbit retina.

RESULTS. In cell cultures, 10–1000 μM glutamate resulted in a dose-dependent loss of neurons, but not of glia. The absence of glutamate toxicity in glia was confirmed in C6 glioma cells. Betaxolol, but not timolol or metoprolol, significantly increased survival (from 52% of control in glutamate-only to 78% with 10 μM betaxolol) after excitotoxic insult. UK14,304 also increased survival (from 62% of control in glutamate only to 100% and 101% of control with 10 and 100 μM UK14,304, respectively). This effect was blocked by the specific α2-antagonist, yohimbine. Both UK14,304 and brimonidine (10–100 μM) reduced glutamate-induced [Ca2+]i increases in retinal neurons in culture. The actions of the α2-agonists in reducing glutamate-induced [Ca2+]i increases, were reduced by yohimbine (1 μM). Betaxolol (100 μM) reduced N-methyl-D-aspartate (NMDA)-induced increases of [Ca2+]i in RGCs in intact retina.

CONCLUSIONS. Betaxolol reduced glutamate excitotoxicity in retinal neurons in vitro through a mechanism independent of β-AR interactions. UK14,304, acting through α2-ARs, was also neuroprotective in vitro. The neuroprotective actions of beta-
these agents protect retinal neurons from glutamate excitotoxicity in vitro by increasing neuronal survival, and if so, to determine whether such neuroprotection effects are associated with receptor-mediated mechanisms and/or decreased calcium influx, as assessed with calcium-imaging in both isolated retinal neurons and in RGCs in an intact retina preparation.

**Materials and Methods**

**Materials**

All culture reagents were obtained from Gibco BRL (Burlington, Ontario, Canada), unless noted otherwise. All other drugs and chemicals were obtained from Sigma Chemical Co. (St. Louis, MO), unless noted otherwise. BDNF and CNTF were provided by Regeneron Pharmaceuticals (Tarrytown, NY), betaxolol by Schwarz Pharma (Boulogne-Billancourt, France), and brimonidine tartrate by Allergan (Irvine, CA). Lysosomal and plasma membrane markers (Zapoglobin) were purchased from Coulter Electronics (Hialeah, FL). The Ca$^{2+}$ indicator dyes fura-2 AM, calcium green-1 dextran, and pluronic acid F-127, were purchased from Molecular Probes (Eugene, OR). In all cases l-glutamate was used but is referred to simply as glutamate.

**Retinal Cell Cultures**

Retinal neurons and glia were isolated from retinas obtained from 6- to 10-day-old Long-Evans rats. All procedures were performed in accordance with the ARVO Statement for the Use of Animals in Ophthalmic and Vision Research and the Dalhousie University Committee for the use of Laboratory Animals. Briefly, rat pups were anesthetized with halothane and decapitated. The eyes were enucleated and posterior eyecups were placed in Hanks’ balanced salt solution (HBSS) and 1% gentamicin. The neural retinas were gently peeled off with fine forceps and incubated at 37°C in 0.02% trypsin for 5 minutes or for 30 minutes in papain (20 U/mL; Worthington Biochemical, Lakewood, NJ). Fetal bovine serum (10%, FBS) was added to the tube to halt the enzyme reaction. Retinal cells were obtained by trituration in growth medium (Neurobasal medium, 10% FBS, 1 ng/mL ciliary neurotrophic factor [CNTF], 10 ng/mL brain-derived neurotrophic factor [BDNF], N$_2$ supplement, and 1% gentamicin). The resultant cell suspension was seeded at a density of 250,000 cells/mL on poly-l-lysine–coated coverslips in 24-well culture plates. Cultures were maintained at 37°C in a humidified atmosphere of 5% CO$_2$/95% air, and half the growth medium was replaced every 2 to 3 days.

The identification of neurons in the mixed neuron-glial cultures was first established by double-label immunocytochemistry with a mouse antibody against neuron-specific β-tubulin (TUJ1; BabCo, Richmond, CA) and contrasted to the number of glial cells identified with a rabbit antibody against glial fibrillary acidic protein (GFAP). Primary antibody labeling was visualized with fluorescent dye conjugated to secondary antibodies (a goat anti-mouse IgG secondary antibody conjugated to Alexa 546 and goat anti-rabbit IgG secondary conjugated to Alexa 488; both from Molecular Probes). Cell density was determined by direct counts of the number of cells in 10 successive 680 × 460-μm fields (camera field of view for 40× objective) in three separate experiments. A C6 glioma cell line (American Type Culture Collection, Manassas, VA) was used for studies requiring a pure CNS glial culture. These cells were maintained in DMEM containing 10% fetal calf serum.

**Cell Survival Assays**

Mixed retinal neurons and glial cultures grown for 7 days in 24-well culture dishes were divided into groups and treated for 24 hours with one of the following: (1) treatment medium (TM) consisting of DMEM, 10% FBS, N$_2$ supplement, 10 ng/mL BDNF, 1 ng/mL CNTF, and gentamicin; (2) TM + glutamate (TMG); or (3) TMG + drug (TMG + D). Cell survival was assayed by a cell lysis method, as previously described. Briefly, the cells were removed from the 24-well plates with trypsin and centrifuged for 5 minutes, and the cell pellet was washed and resuspended in 300 μL of a 1:10 dilution of lysis solution. Viable cell nuclei were counted with a hemocytometer. Mean cell counts were obtained from three to four wells per treatment group (TM, TMG, or TMG + D) and expressed as a percentage of the number of viable cells in the control TM group. Mean percentage data ± SEM were obtained from three to six separate experiments. The drugs used were betaxolol, metoprolol, and timolol (β-blockers) and UK14,304 and brimonidine (α$_2$-agonists), and the effect of each was tested over the range of concentrations from 1 to 1000 μM. The specificity of the effect of UK14,304 was further investigated with the α$_2$-AR antagonist yohimbine at 1 and 10 μM. Solutions of UK14,304 were prepared from stocks dissolved in dimethyl sulfoxide (DMSO). Therefore, in experiments in which UK14,304 was used, equivalent amounts of DMSO were also added to the other treatment solutions (TM, TMG).

Differences between mean data were compared for statistical significance with a two-tailed t-test or one-way analysis of variance (ANOVA) and the Dunnett multiple comparison test (Instat; GraphPad, San Diego, CA).

**Calcium Imaging of Retinal Neuron Responses to Glutamate**

Isolated rat retinal neurons were maintained in serum-free Neurobasal medium with 1 mM glutamine, 10 μg/mL gentamicin, 2% B27 supplement, 40 ng/mL BDNF, and 10 μM forskolin. The neurons were loaded with the ratiometric calcium-indicator dye fura-2. Cells were incubated in 5 μM fura-2-acetoxyxymethyl ester (AM) dissolved in modified HBSS (described later) for 20 minutes in the dark at 37°C. The fura-2 AM was first dissolved in DMSO (0.1% final concentration in HBSS) and then solubilized in HBSS containing 0.1% pluronic acid F-127. After loading, the cells were washed in HBSS for at least 15 minutes before imaging.

To assess the effect of glutamate on [Ca$^{2+}$], we used a modified Mg$^{2+}$-free HBSS (pH 7.4, 20 mM HEPES, 1.5 mM CaCl$_2$). All treatment solutions containing glutamate, UK14,304, and yohimbine were prepared from stocks and dissolved in the modified HBSS. The fura-2-loaded neurons plated on coverslips were maintained in a chamber (ΔTC3; Bioptechs, Butler, PA) that was constantly superfused with HBSS at 34°C to 36°C (SH-27 Inline heater, TC-324B controller; Warner Instruments, Hamden, CT) and bubbled with 100% oxygen. Solutions were delivered to the chamber by a peristaltic pump (Gilson, Middleton, WI) at a rate of 2 mL/min. During treatments, image pairs were collected as often as every 3 seconds, but to limit photodamage, images were collected less frequently (20 seconds) during intervening periods.

The neurons loaded with fura-2 were imaged with a cooled charge-coupled device (CCD) camera (Sensicam; PCO Computer Optic, Kelheim, Germany) fitted to a microscope (Axioskop FS; Carl Zeiss, Oberkochen, Germany) using a water-immersion objective (numeric aperture 0.80; Achroplan 40×; Zeiss). Fura-2 fluorescence was produced by excitation from a 75W xenon lamp (Ludl Electronic Products, Hawthorne, NY) and appropriate filters (XPO4 set, excitation 340 or 380 nm; emission 510 nm; dichroic >430 nm; Omega Optical, Brattleboro, VT). To reduce photodamage and photobleaching, excitation illumination was filtered with a ~0.5 log neutral density filter and 610 × 512-pixel or 320 × 256-pixel images acquired. The duration of illumination was limited (400 msec) by an electronic shutter (Uniblitz, Rochester NY). Images at 340 and 380 nm excitation were captured and converted to ratiometric (340/380 nm) images by an imaging system (Imaging Workbench 2.2, Axon Instruments, Foster City, CA) and saved to the hard disk of a computer. The mean ratio over a large area of a neuron, well separated from the edge of the cell, was measured for each captured ratiometric image.

Free [Ca$^{2+}$], was elevated in cultured cells by application of a short pulse (20 seconds) of 10 μM glutamate in HBSS. This protocol was chosen because it resulted in reproducible increases of [Ca$^{2+}$]$_i$ within the dynamic range (below saturation) of fura-2 fluorescence. The retinal neurons were treated with UK14,304 (10 – 100 μM) or brimonidine (100 μM) for 5 minutes before and 3 minutes after the glutamate burst. Solutions of UK14,304 were prepared from stocks...
dissolved in DMSO. Therefore, in experiments in which UK14,304 was used, equivalent amounts of DMSO were also added to the other treatment solutions. For experiments with yohimbine, the neurons were pretreated first for 5 to 10 minutes with 1 μM yohimbine alone and then with yohimbine and UK14,304 together for the same period as in the experiments with UK14,304 alone. The time between consecutive bursts of glutamate was kept at 10 minutes to allow sufficient time for [Ca²⁺]i levels to recover to a baseline level.

The peak fura-2 ratios were normalized to the initial treatment with glutamate and differences between the peak fura-2 ratios (pretreatment, in presence of UK14,304 or brimonidine, and recovery) were tested for statistical significance with the Freidman repeated measures ANOVA followed by a modified Student-Newman-Keuls multiple-comparison test. The differences between the peak ratios for UK14,304 alone versus UK14,304+yohimbine were tested for statistical significance with the Mann-Whitney rank sum test. All analyses were performed on computer (SigmaStat software; SPSS Science, Chicago, IL).

**Calcium Imaging in an Intact Rabbit Retina Preparation**

Calcium imaging of RGCs in intact retina was performed by a method described previously. Rabbits were used for these experiments, because the isolated retina preparation from this animal is robust and well characterized and is the preparation in which the calcium-imaging technique used was developed. In brief, adult rabbits were killed by barbiturate overdose and both eyes enucleated. The anterior segment and vitreous were removed, and the posterior segment immersed in Ames medium, buffered to pH 7.4 with HEPES, and bubbled continuously with 100% oxygen at room temperature. The retina was dissected from the eyebuc, placed on a glass microscope slide, and cut into several pieces with a scalpel blade. The pieces of retina were then mounted on a filter (Millipore, Bedford, MA) with the ganglion cell layer uppermost.

Using a 10-μL syringe fitted with a sharpened tapered 26-gauge needle (Hamilton, Reno, NV) a small amount (<1 μL) of 10% (wt/vol) calcium green-1 conjugated to 10,000 MW dextran (CaGD) and dissolved in purified water was deposited within the substance of the retina. After application of the dye, retinas were incubated in the dark for at least 2 hours at room temperature in Ames medium bubbled continuously with 100% oxygen.

As described previously, dextran-conjugated dye injected into the retina labels a variety of neuronal cell types near the injection site, including amacrine cells displaced to the ganglion cell layer, but it also labels RGC axons that pass through the injection site. In the latter case, the dextran is then retrogradely transported to groups of RGC somata up to 800 μm from the injection site. In the present work, only the RGCs retrogradely labeled by CaGD were studied.

Retinal wholemounts mounted on a filter were transferred to the same imaging setup as that used in the studies of isolated cells, and the conduct of experiments was similar to that described for the studies of isolated cells. Because CaGD is not a ratiometric dye, fluorescence was imaged using only a single filter set (XF100; Omega Optical) and the data reported as increases of fluorescence intensity divided by the resting intensity (ΔF/F). Free [Ca²⁺]i, within RGCs was increased by application of 50 μM N-methyl-D-aspartate (NMDA). The effect of betaxolol (100 μM) on NMDA-induced increases in [Ca²⁺]i, was assessed by applying betaxolol to the isolated retina both before (8 minutes) and during the treatment with NMDA. Differences in CaGD ΔF/F during different treatments were tested for statistical significance with a paired t-test of the raw data (Statview SE; Abacus Concepts, Berkeley, CA).

**RESULTS**

**Immunocytochemical Identification of Neurons and Glia In Vitro**

The identification of neurons in the mixed neuron-glia cultures was first established by using double-label immunocytochemistry with antibodies against neuron-specific β-tubulin and GFAP (Fig. 1). The neuron-specific, β-tubulin–labeled cells (neurons) had compact cell bodies and processes longer than twice the size of the somata, whereas GFAP labeling was restricted to cells (glia) with flattened morphology and processes less than twice the size of the somata. Colocalization was never observed. Therefore, in subsequent survival and calcium-imaging studies, cells were classified as neurons if they possessed the appropriate morphology together with processes (neurites) greater than twice the size of the somata. Cells without these long processes were considered to be glia. On average, neurons constituted approximately 35% to 40% of the total population of cultured cells.

**Effect of Glutamate on Retinal Neuron Survival**

The effects of increasing concentrations of glutamate on neuronal survival were examined in mixed rat retinal neuron-glial cultures. Addition of glutamate at concentrations of 1 to 1000 μM to mixed retinal cell cultures for 24 hours resulted in a significant dose-dependent decrease in the number of viable cells from 57% ± 2% of the control group at a dose of 1 μM glutamate, to 51% ± 1%, 37% ± 1%, and 19% ± 0.5% of the control group at 10, 100, and 1000 μM glutamate, respectively (P < 0.05; Fig. 2A). In subsequent experiments, a dose of 100 μM glutamate was used in cell survival assays, because this was the dose in mixed retinal cell cultures that consistently produced approximately 50% or greater cell loss. The cell loss in mixed cultures exposed to glutamate consisted primarily of neurons (Fig. 2B). After 24 hours’ exposure to glutamate, there was a 91% decrease (P < 0.01) in the number of neurons, defined by the presence of neurites, but only a 14% decrease (P > 0.50) in the number of glial cells, defined by absence of neurites, flattened morphology, and positive staining for GFAP (Fig. 2B). Figure 2C shows the response of a pure glial cell line (CG glioma) to glutamate treatment. These data demonstrate that 10 and 100 μM glutamate had no significant (P > 0.40) effect on glial cell survival.

**Effects of Betaxolol, Timolol, and Metoprolol on Retinal Neuron Survival**

Figure 3 shows data from experiments examining the survival of retinal cultures in the presence of 100 μM glutamate alone.
Neuroprotective Effects of Adrenoceptor Drugs In Vitro

Glutamate (10 μM) increased the fura-2 fluorescence ratio from a resting level of approximately 0.5 (Fig. 5A) to a peak of approximately 1.1 during exposure to 10 μM glutamate (Fig. 5B). One hundred twelve cells were studied and, on average, 10 μM glutamate produced a peak ratio of 1.07 ± 0.01 (mean ± SEM), increasing from an average resting ratio of 0.51 ± 0.01.

In the presence of 10 or 100 μM UK14,304, there was a significant decrease in the peak ratio reached by exposure to 10 μM glutamate. For example, in the cell shown in Figure 6A the presence of 100 μM UK14,304 reduced the glutamate-induced peak fura-2 ratio to 0.997, an 11% decrease from the control response of 1.12. On average, UK14,304 reduced sig-

Effect of UK14,304 on Retinal Neuron Survival

In the groups treated with a dose of 10 or 100 μM UK14,304 plus glutamate (Fig. 4A) there was a significant increase (P < 0.05) in cell survival (109%) from 0.51 (mean approximately 1.1 during exposure to 10 μM glutamate) to a peak of approximately 1.12. On average, UK14,304 reduced significantly the number of neurite-bearing (neurons) and/or glia. Cell viability was determined from the number of intact nuclei after cell lysis. Data are the mean ± SEM of results derived from three to four separate experiments. (A) *P < 0.05 for the number of viable cells in TMG groups compared with the TM-only group. (B) **P < 0.01 for neurite-bearing cells in TMG (100 μM) compared with the number of neurite-bearing cells in control TM only (0).

Effect of α2-AR Activation on the Glutamate-Stimulated [Ca2+]i Increase in Isolated Rat Retinal Neurons

Glutamate (10 μM) plus various concentrations (1-1000 μM) of the β1-selective AR blockers betaxolol and metoprolol or the non-selective β-AR blocker timolol. Betaxolol produced a dose-dependent increase in viable cells up to 100 μM, compared with the glutamate-only treatment group (Fig. 3A). Viable cells (mean ± SEM) in the presence of 1, 10, 100, and 1000 μM betaxolol were 56% ± 10%, 78% ± 2%, 93% ± 14%, and 86% ± 50% of control glutamate-free group compared with 53% ± 4% of control in the presence of glutamate alone. This increase in cell survival was significant at 10 μM betaxolol (P < 0.01) with increased variability at higher doses limiting the significance (P > 0.05 and P > 0.30 at 100 and 1000 μM, respectively) of the effect of betaxolol. In contrast to betaxolol, the nonspecific β-AR blocker timolol failed (P > 0.50) to increase cell survival at all doses tested (1-1000 μM) over that observed in the glutamate-only treatment group (Fig. 3B). We also tested another β1-selective AR blocker, metoprolol, at 1 to 1000 μM (Fig. 3C). As with timolol, the number of cells surviving in the metoprolol-treated groups was not significantly different (P > 0.50) from that in the glutamate-only group at all doses tested.

Stimulated [Ca2+]i

Stimulated [Ca2+]i of the presence of 100 μM yohimbine (Fig. 4B), the number of surviving cells was not significantly different from that in the glutamate-only treatment group, but was significantly different from the glutamate plus α2-AR agonist-treated group (P < 0.05). This suggests that in contrast to the β-AR blocker betaxolol, the α2-AR agonist UK14,304 exerts a neuroprotective effect on retinal neurons through interacting with α2-AR on retinal neurons and/or glia.

Glutamate-induced excitotoxic retinal neuron death. (A) Number of viable cells after 24 hours of glutamate treatment, as assessed by counting the density of intact nuclei after cell lysis. Mixed retinal neuron cultures were plated at a density of 250,000 cells/ml and treated with TM plus increasing doses of glutamate (TMG, 1-1000 μM). Control cultures were treated with TM without glutamate (TM, 0). (B) Morphologic analysis of the number of neurite-bearing (neurite-bearing cells) and non-neurite-bearing (non-neurite-bearing cells) in 10 successive 680 μM high-power (40X objective) fields, before and after 24 hours of glutamate (10 μM) treatment (n = 3). (C) C6 glioma rat cell line cultures treated for 24 hours with either 10 or 100 μM TM plus glutamate or TM without glutamate. Cell viability was determined from the number of intact nuclei after cell lysis. Data are the mean ± SEM of results derived from three to four separate experiments. (A) *P < 0.05 for the number of viable cells in TMG groups compared with the TM-only group. (B) **P < 0.01 for neurite-bearing cells in TMG (100 μM) compared with the number of neurite-bearing cells in control TM only (0).
significantly ($P < 0.01$) the effect of 10 μM glutamate on the peak fura-2 ratio with a dose-dependent decrease of 7% ± 1% ($n = 21$) and 15% ± 1% ($n = 31$) for 10 and 100 μM UK14,304, respectively (Fig. 6D). In the example shown in Figure 6A, there was essentially complete recovery of the
response to 10 μM glutamate after washout of UK14,304. On average, however, recovery was incomplete (Fig. 6D). Although the response to glutamate after washout of UK14,304 remained significantly different from the response produced before treatment with UK14,304 ($P < 0.01$) the peak ratio was significantly increased compared with that achieved during the treatment ($P < 0.01$). Similar results were achieved when brimonidine, an aqueous-soluble analogue of UK14,304,
was used (Fig. 7). The glutamate response was reduced by 17% \( \pm 3\% \) (\( n = 5 \)) in the presence of 100 \( \mu M \) brimonidine and showed some recovery (10% \( \pm 4\% \) less than control response) after washout. The lack of full recovery was not due to a reduction of the response to glutamate, because multiple treatments with glutamate in the absence of UK14,304 or brimonidine produced similar ratio increases (data not shown, but see Fig. 6B). In some cells, such as the cell illustrated in Figure 7A, UK14,304 or brimonidine decreased the resting fura-2 ratio, but on average the decrease was not statistically significant.

To determine whether the effect of UK14,304 on glutamate-induced elevations of \( [\text{Ca}^{2+}]_i \) were due to the specific action at \( \alpha_2 \)-ARs, the specific antagonist yohimbine was used. Initial experiments showed that 1 \( \mu M \) yohimbine alone had no effect (\( n = 20; \ P > 0.05 \)) on the glutamate-induced increase of

![Figure 7](http://iovs.arvojournals.org/pdfaccess.ashx?url=/data/journals/iovs/932916/)

**FIGURE 7.** Brimonidine limits the increase of \( [\text{Ca}^{2+}]_i \) in isolated rat retinal neurons. (A) Effect of 100 \( \mu M \) brimonidine on the peak of the fura-2 fluorescence ratio produced by 10 \( \mu M \) glutamate. (B) Mean data for all cells studied (\( n = 5 \)) normalized to the first exposure (pretreatment) to 10 \( \mu M \) glutamate. \( **P < 0.01 \) compared with glutamate pretreatment; \( \dagger P < 0.01 \) recovery after pretreatment with brimonidine + glutamate.

![Figure 8](http://iovs.arvojournals.org/pdfaccess.ashx?url=/data/journals/iovs/932916/)

**FIGURE 8.** Effect of betaxolol on the increase of \( [\text{Ca}^{2+}]_i \) in RGCs in intact rabbit retina. (A) Change in fluorescence produced by 50 \( \mu M \) NMDA, in the presence and after washout of 100 \( \mu M \) betaxolol. (B) Mean data from 11 cells normalized to the response produced by NMDA. \( *P < 0.05 \).
[Ca^{2+}], (Fig. 6B, 6D). These experiments also demonstrated that the reduced peak ratio during and after UK14,304 treatment was not simply due to a reduction of the response of the cells to glutamate over time or a result of repeated glutamate treatments. Yohimbine (1 μM) limited the effect of UK14,304 on glutamate-induced increases of [Ca^{2+}]. This is illustrated graphically for a single cell in Figure 6C, where the effect of 100 μM UK14,304 was essentially abolished by yohimbine. On average (Fig. 6D), 1 μM yohimbine reduced but did not completely eliminate the effect of UK14,304. Although glutamate responses in the presence of both yohimbine and UK14,304 did not reach control (pretreatment) levels (P < 0.05), the peak ratio obtained was greater than in the presence of UK14,304 alone and this was statistically significant (n = 20, Kruskal-Wallis; P < 0.01; Fig. 6D).

Effect of Betaxolol on NMDA-Induced Increases of Rabbit RGCs [Ca^{2+}] in Intact Retina

RGCs within the ganglion cell layer of the rabbit retina were loaded with CaGD (Fig. 5C) and treated with 50 μM NMDA (Fig. 5D). NMDA was used instead of glutamate, because the uptake of glutamate by neurons and glia in intact retina preparation severely limits the level of extracellular glutamate, even if applied exogenously.4 4 Although there was also some loading of axons with CaGD (Fig. 5C), NMDA treatment did not increase fluorescence within axons. Treatment with 100 μM betaxolol reduced the increase of CaGD fluorescence in RGCs produced by 50 μM NMDA (Fig. 8). The protocol used tested the response to 50 μM NMDA first in the presence of 100 μM betaxolol and then after approximately 10 minutes’ washout. As described previously,4 0 this protocol limits reduction of responses that can occur with multiple increases of [Ca^{2+}] in this preparation. An optical recording from a single RGC within the rabbit ganglion cell layer is shown graphically in Figure 8A. The tissue was treated with 100 μM betaxolol for 10 minutes, but at 7 minutes was also exposed to a 2-minute pulse of 50 μM NMDA. In the cell illustrated (Fig. 8A) this resulted in an increase of CaGD fluorescence (ΔF/F) of less than 0.2. After approximately 10 minutes’ washout the tissue was treated again with 50 μM NMDA, and the response was much larger, exceeding a ΔF/F of 0.6. Average results from 11 RGCs are shown in Figure 8B. The responses have been normalized to the control (50 μM NMDA alone) treatment and show that, in the presence of 100 μM betaxolol, there was an approximate 40% decrease (P < 0.05) of the response of the cell to 50 μM NMDA.

DISCUSSION

There is now increasing evidence that both the β2-AR antagonist betaxolol and α2-AR agonists, such as brimonidine, decrease RGC cell death subsequent to increased intraocular pressure, retinal ischemia, or optic nerve crush.5 7–38 However, all these studies were performed in vivo, and drugs were applied topically or systemically, making it impossible to determine whether the action of the drugs was directly neuroprotective or indirect, affecting local blood supply, inflammatory mediators or other nonretinal ocular tissues. Although no studies have assessed the effect of AR drugs on retinal neuron survival in vitro, a few studies have examined the effects of such drugs on ion channels but, for the most part, in neurons isolated from lower vertebrate retinas.3 4,4 5,4 6

Our experiments used a mammalian (rat) in vitro cell culture model of retinal excitotoxicity to examine the neuroprotective actions of the β-AR blockers, betaxolol, metoprolol, and timolol and the α2-AR agonist UK14,304. This in vitro model allowed the actions of the drugs to be studied directly on retinal neurons and glia in isolation of vascular actions and systemic effects. In our mixed retinal cultures, exposure to exogenous glutamate (10–100 μM) produced an approximate 40% to 50% reduction in the number of cells during a 24-hour period. Immunocytochemical and morphologic identification determined that most of the cell death was neuronal. Glial cells in mixed cultures were relatively resistant to such doses of glutamate, and cells from a purified glial cell line (C6 glioma) were also resistant to glutamate over the same range of concentrations (10–100 μM).

Of the β-blockers tested, only betaxolol increased cell survival after excitotoxic insult in primary rat retinal cultures, whereas the nonspecific β-blocker timolol and another β1-AR-selective blocker metoprolol did not. These findings seem to demonstrate that the neuroprotective effect of betaxolol on retinal neurons is independent of β-AR interactions. The α2-agonist UK14,304 was also able to increase retinal cell survival in mixed retinal cultures after excitotoxic stress. This effect was both dose-dependent and receptor-mediated, because it was blocked by the α2-AR antagonist, yohimbine. The decrease in neuroprotection at higher doses of the α2-AR agonist may be associated with agonist-mediated receptor desensitization.4 5

An examination of the effect of betaxolol on NMDA-induced increases of [Ca^{2+}], in RGCs in isolated intact rabbit retina supports and extends previous results in isolated rat RGCs,4 5 showing that betaxolol inhibits glutamate-receptor stimulated increases of [Ca^{2+}], in RGCs in situ. The use of a different species and a different preparation strengthens the view that betaxolol is capable of limiting excitotoxicity arising from excessive or prolonged Ca^{2+} influx. Glutamate-induced [Ca^{2+}] increases in isolated retinal neurons were also decreased by α2-AR agonists at doses comparable to those found to be neuroprotection in vitro. Therefore, some component of the neuroprotection afforded by betaxolol and α2-agonists in vitro and in vivo may be due to decreased Ca^{2+} influx. A decrease in Ca^{2+} influx would be expected to increase cell survival after excitotoxic stress by reducing activation of Ca^{2+}-dependent proteases and limiting free radical production and DNA damage.1 0,1 1,1 5

It has been demonstrated that certain β-AR blockers have additional properties unrelated to their actions at ARs that include actions at other receptors and ion channels. In the case of betaxolol but not other β-AR blockers such as propranolol, this includes block of voltage-dependent Ca channels in peripheral3 8 and retinal vasculature4 7,4 8 and inhibition of neuronal ion channels, Ca^{2+} influx, and neuronal excitability.4 3,4 7,4 9,5 0 In our studies, metoprolol, a β1-AR blocker, failed to increase neuronal survival significantly at a concentration at which betaxolol had significant neuroprotective actions, and timolol, a nonselective β-AR blocker that has been shown not to have Ca channel-blocking action in cultured RGCs,4 5 was also ineffective at increasing retinal neuron survival in primary cultures. These data support the hypothesis that in the in vitro retinal cell models used, the neuroprotective actions of betaxolol can be primarily attributed to its direct suppressive actions on neuronal ion channels and Ca^{2+} influx.

In contrast to the results obtained with betaxolol, our data suggest that the neuroprotective effects of UK14,304 on retinal neurons were mediated by a specific action at α2-ARs, in particular because the neuroprotection was blocked by the α2-AR antagonist yohimbine. These data are consistent with evidence that the α2-AR agonists brimonidine and clonidine are neuroprotective in animal models of ischemia, in pressure-induced retinal hypoxia, and after mechanical damage to RGC axons.5 0,5 1,5 2,5 1,5 2 The mechanism(s) for this in vivo neuroprotection also appears to involve α2-AR activation, because selective antagonists
such as rauwolscine or yohimbine block the neuroprotective action that may indicate
L-type Ca$^{2+}$-channels and/or release of Ca$^{2+}$-influx by the enhancement of basic
2-agonists clonidine and xylazine activates extracellular signal-regulated kinases p42/p44 primarily in Müller cells,\textsuperscript{55} an action that may indicate 2-AR-mediated regulation of the release of neuroactive factors from Müller cells that could influence retinal neuronal survival. Activation of neuronal 2-ARs may also act directly to decrease Ca$^{2+}$ influx and excitation transmitter release.\textsuperscript{56} Radioligand binding studies and cloning have demonstrated the presence of several 2-AR isoforms (2A2, 2C2, and 2D2) in the inner plexiform layer of the mammalian retina.\textsuperscript{56}–\textsuperscript{58} In many types of neurons, 2-ARs are coupled to voltage-dependent N-type and P/Q-type Ca channels through the G$\alpha_1$i family of proteins to inhibit Ca$^{2+}$ current, Ca$^{2+}$ influx, and neurotransmitter release.\textsuperscript{59}–\textsuperscript{61} RGCs and retinal neurons are known to express different Ca channel subtypes, including a toxin-resistant Ca$^{2+}$ current component and N-type and L-type Ca$^{2+}$ currents.\textsuperscript{62,63} Therefore, activation of 2-ARs may result in inhibition of Ca channels on retinal neurons. Another laboratory has examined binding of the 2-AR agonist clonidine to voltage-dependent Ca and Na channels as well as the effects of clonidine on radiolabeled Ca$^{2+}$ influx in intact isolated rat retina. Their studies have reported that, unlike betaxolol, clonidine did not directly interact with voltage-dependent L-type Ca channels or Na channels and did not produce a significant decrease in Ca$^{2+}$ influx.\textsuperscript{52,64}

Our data examining alterations of [Ca$^{2+}$]i in isolated retinal neurons indicate that both UK14,304 and brimonidine can increase survival of isolated retinal neurons in vitro confirming that these drugs can act as neuroprotectants through actions independent of vascular targets. The effective concentrations of drugs used in our studies of isolated cells and retina (1-100 $\mu$M) and others using isolated tissue preparations\textsuperscript{34,49} are in the clinically relevant range, estimated to be approximately 1 $\mu$M in plasma, with greater drug levels accumulating during prolonged administration.\textsuperscript{48,65} In vivo experiments in rats and monkeys using topical and systemic application of $\beta$-blockers indicate that levels of these drugs with repeated administration accumulate in the micromolar range in the retina and optic nerve head, with even greater levels present in the choroid, sclera, and iris ciliary body.\textsuperscript{56,67} Our results comparing different $\beta$-AR antagonists support and extend previous data by demonstrating that the neuroprotective properties of betaxolol in the retina are due, at least in part, to a direct effect on neurons, are independent of specific $\beta$-AR receptor interactions and probably arise from the actions of betaxolol on neuronal ion channels and [Ca$^{2+}$]i.\textsuperscript{34,43,49,50,52} In contrast to betaxolol, the survival-promoting actions of the 2-AR agonist UK14,304 occurred through the activation of 2-AR receptors. However, our data demonstrating that 2-AR agonists can modulate [Ca$^{2+}$]i after exposure to glutamate suggest that in addition to other reported actions, such as increasing retinal levels of bFGF,\textsuperscript{28,52} they may also act directly on retinal neuron 2-AR receptors to limit increases in [Ca$^{2+}$]i.

References


