Slow-Releasing Tranilast in Polytetrafluoroethylene/Polylactide-co-glycolide Laminate Delays Adjustment after Strabismus Surgery in Rabbit Model

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PURPOSE. To determine the usefulness of slow-releasing tranilast in polytetrafluoroethylene/polylactide-co-glycolide (PTFE/PLGA) laminate for delayed adjustable strabismus surgery.

METHODS. A prospective, masked-observer, controlled study was performed in 25 rabbits. Fifty rabbit eyes were divided randomly into three groups. After a recession of the superior rectus muscle (SRM), a PTFE/PLGA laminate containing tranilast, PTFE alone, or balanced salt solution was applied beneath and over the SRM in the PTFE/PLGA-tranilast group (group P-T), the PTFE group (group P), and the control group, respectively. Delayed adjustment was performed by a masked observer once on each SRM 3 or 5 weeks after surgery. Adjustability, adjustment lengths, forces required, and adhesion degrees were evaluated.

RESULTS. In the control group, adjustment was impossible in any eye 3 or 5 weeks after surgery. In group P, adjustment was possible in 5 of 8 eyes 3 weeks after surgery and in 5 of 10 eyes 5 weeks after surgery. In group P-T, adjustment was possible in 8 of 10 eyes 3 and 5 weeks after surgery. On comparing adjustability, a significant difference was observed between group P-T and the control group 3 and 5 weeks after surgery (P = 0.006, P = 0.006, respectively). A significant difference was observed between group P-T and the control group (P = 0.016) in terms of adhesion between SRMs and conjunctivae 5 weeks after surgery.

CONCLUSIONS. Slow-releasing tranilast in PTFE/PLGA was found to reduce adhesion and allowed delayed adjustment in most eyes for up to 5 weeks after surgery. (Invest Ophthalmol Vis Sci. 2007;48:699–704) DOI:10.1167/iovs.06-0775

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bismus surgery. In this experimental study, we evaluated the effectiveness of PTFE/PLGA film containing tranilast to facilitate delayed adjustment after strabismus surgery in a rabbit model.

**Materials and Methods**

All experiments were conducted in accordance with the ARVO Statement for the Use of Animals in Ophthalmic and Vision Research. The protocol was approved by the Institutional Animal Care and Use Committee of Seoul National University Hospital.

Twenty-five New Zealand White rabbits, each weighing 2.0 to 3.0 kg, underwent 5-mm recession of the superior rectus muscles (SRMs) with the use of double-armed, nonabsorbable 5-0 polyester (5-0 Ethibond; Ethicon, Somerville, NJ) sutures and were subsequently adjusted 3 and 5 weeks after surgery. PTFE/PLGA containing sustained-release tranilast was used as a physical barrier in group P-T, whereas PTFE film containing tranilast was used in group P. In the control group (group C), no physical barrier was used. Fifty SRMs were divided among three experimental groups as follows: group C (10 muscles; 5 for examination at 3 weeks and 5 at 5 weeks), group P (20 muscles; 10 for examination at 3 weeks and 10 at 5 weeks), and group P-T (20 muscles; 10 for examination at 3 weeks and 10 at 5 weeks). Two eyes of each rabbit were randomly assigned to three groups to minimize inter-rabbit variation.

**Procedures**

General anesthesia was induced by administering 30 to 45 mg/kg ketamine hydrochloride and 5 to 10 mg/kg xylazine hydrochloride intramuscularly, and topical anesthesia was achieved using proparacaine hydrochloride. Muscle recession procedures with or without a physical barrier were performed as previously described.18

Polyvinylpyrrolidone-iodine was applied to the eyelids for preoperative surgical antisepsis. Limbal peritomy was performed from 10 o’clock to 2 o’clock. The SRM was isolated on a Jameson hook, and intermuscular connections were dissected. The superior oblique tendon was disinserted and allowed to retract from the surgical field. The SRM was then placed on a double-armed 5-0 polyester (5-0 Ethibond; Ethicon) suture close to the insertion and disinserted from the globe. In group P-T (20 SRMs), a separate sheet (1 × 2 cm) of PTFE/PLGA containing tranilast was placed between the sclera and the SRM and folded to separate the conjunctiva and the SRM. An identical process was used in the PTFE without tranilast group (group P; 20 SRMs), but no barrier was inserted in the control group (10 SRMs). Finally, the SRM was recessed 5 mm and reattached to the original insertion using double-armed, nonabsorbable 5-0 polyester (5-0 Ethibond) suture. A dial tension gauge (DT-50; Castroviejo, UK) was used to measure the force required for adjustment. The suture was attached to the SRM with a Castroviejo caliper, and the force required was measured with the gauge.

**Delayed Adjustment**

In a masked random fashion, the SRMs were adjusted under anesthesia 3 or 5 weeks after surgery in all three groups. The delayed adjustment procedure was performed as follows. The previous conjunctival incision site was opened with the tip of a curved needle holder or scissors. After the SRM had been fully exposed, a PTFE or a PTFE/PLGA membrane was removed before adjustment. A dial tension gauge (DT-50; Teclock Inc., Okaya, Japan) was used to control the amount of force applied. The noose suture attached to the SRM was hooked with the bar of a dial tension gauge. The SRM was then moved anteriorly as much as possible. This adjustable distance was measured with a Castroviejo caliper, and the force required was measured with the gauge.

**Preparation of Polymeric Film Fabrication**

Poly(lactide-co-glycolide) 50:50 (PLGA; average molecular weight, 40,000–75,000; Sigma, St. Louis, MO) containing tranilast (Choongwae Pharma Corp., Seoul, Korea) was cast onto PTFE film with a solvent casting technique. Brieﬂy, tranilast (144 mg) and PLGA (1000 mg) were dissolved in 10 mL carbonate ﬁber (Fisher Scientiﬁc, Suwanee, GA) with stirring for 30 minutes at room temperature. Blank and drug-containing PLGA solutions were cast onto glass plates and onto PTFE ﬁlm-coated glass plates. Cast polymer ﬁlms were dried in a refrigerator for 48 hours to allow the acetone to evaporate slowly and then were dried for 24 hours at room temperature. PLGA ﬁlms and PTFE ﬁlms coated with PLGA were peeled from the glass plates and cut into pieces measuring 1 cm × 1 cm. Drug loadings on ﬁlm weighed approximately 1.0 mg/cm².

**Determination of Tranilast Loading PLGA**

Polymeric films were dissolved in 2 mL acetonitrile and vortexed for 1 minute. The polymer solutions obtained were then ﬁltered through a 0.45-μm membrane ﬁlter to remove precipitated material; Acetonitrile (4.5 mL) was then added to 0.5 mL of ﬁltrate and vortexed for 30 seconds. One hundred microliters of this solution was diluted to 900 μL with mobile phase, and a 10-μL aliquot was directly injected into a high-performance liquid chromatograph. Tranilast loadings were then determined as previously described.21

The high-performance liquid chromatography (HPLC) system comprised a separation module (2690; Waters, Milford, MA) with pump, autosampler, UV detector (2487; Waters), and C18 reversed-phase column (Capcellpak MGII, 5 μm, 250 mm × 1.5 mm; Shiseido, Tokyo, Japan). Data were analyzed (MassLynx software; Micromass, Manchester, UK). The mobile phase consisted of 40% (vol/vol) acetonitrile and 60% (vol/vol) 10 mM phosphate buffer (pH 3.5). The volume injected was 10 μL, and the flow rate used was 0.10 mL/min. Under these conditions, linear calibration curves of tranilast were obtained in the concentration range 0.5 to 8 μg/mL (r² > 0.99).

**In Vitro Release of Tranilast**

Tranilast release from the polymer laminate was studied in phosphate-buffered saline (PBS). A polymer film sample (1 cm × 1 cm) was placed in 23 mL PBS (pH 7.4) containing 0.1% sodium azide as a preservative, and magnetically stirred in a circulating water bath 37°C. At predetermined times (0, 0.17, 0.25, 0.33, 0.42, 1, 2, 3, 5, 6, 7, 10, 15, 17, 21, 27, and 30 days), 400-μL aliquots of the medium were collected, replaced with an equal volume of fresh PBS, and filtered through a 0.45-μm membrane ﬁlter. Ten microliters of this solution was directly injected to the HPLC for tranilast quantification. In vitro release experiments were repeated three times to obtain a release proﬁle of tranilast.

**Evaluation of Adhesions**

At the time of adjustment, adhesions between muscle, PTFE or PTFE/PLGA membrane, sclera, and conjunctiva were evaluated and recorded. Adhesions were classiﬁed as SRM/C (superior rectus muscle/conjunctiva) or as SRM/S (superior rectus muscle/sclera) when they were, respectively, located above or below an SRM. Adhesion severities were scored from 0 to 4 according to criteria described in previous report,18 as follows: 0 = no adhesion; 1 = milky adhesion easily separable with blunt dissection; 2 = mild to moderate adhesion with freely dissectible plane; 3 = moderate to dense adhesion with difﬁcult dissection; and 4 = nondissectible plane. Animals were killed after delayed suture adjustment by administering a 10-mL intravenous injection of sodium pentothal.

**Postmortem Histologic Examination**

Histopathologic studies were performed to evaluate inﬂammation and ﬁbrosis between sclera, SRM, and conjunctiva. Tissues in each of three groups were examined macroscopically and microscopically using hematoxylin and eosin staining and Masson trichrome staining.
nation of drug diffusion through the PLGA film and hydrolytic degradation of PLGA.

**Adjustability**

In the control group, adjustment was impossible in any eye 3 and 5 weeks after surgery. In group P, adjustment was possible in 5 of 8 eyes 3 weeks after surgery and in 5 of 10 eyes at 5 weeks after surgery. In group P-T, adjustment was possible in 8 of 10 eyes at 3 weeks and 5 weeks after surgery (Table 1).

On comparing adjustability 3 and 5 weeks after surgery, a significant difference was found among three groups ($P = 0.013, P = 0.015$, respectively). Subsequent pairwise comparisons showed that the difference between group P-T and the control group was significant ($P = 0.006, P = 0.006$, respectively; Bonferroni corrected). No significant difference was observed between groups P and P-T or between group P and the control group 3 weeks ($P = 0.608, P = 0.075$, respectively) or 5 weeks after surgery ($P = 0.349, P = 0.101$, respectively).

**Adjustment Amount and Force Required**

In group P, average adjustment (advancement) amount and force were 2.10 mm and 54.00 g 3 weeks after surgery and 2.35 mm and 50.80 g 5 weeks after surgery. In group P-T, the average adjustment amount and force were 2.31 mm and 34.12 g 3 weeks and 2.43 mm and 42.75 g 5 weeks after surgery (Table 1). On comparing adjust amount, no difference was found between groups P and P-T at 3 or 5 weeks after surgery ($P = 0.945, P = 0.724$, respectively). For adjust forces, a significant difference was found between groups P and P-T at 3 weeks after surgery ($P = 0.030$) but not at 5 weeks after surgery ($P = 0.524$).

**Degree of Adhesion between SRM and Conjunctiva**

A significant difference was observed among three groups with respect to the degree of adhesion between SRMs and conjunctiva 3 or 5 weeks after surgery ($P = 0.002, P = 0.023$, respectively). Pairwise comparisons revealed significant differences between groups P and P-T ($P = 0.001$; Bonferroni corrected) 3 weeks after surgery and between group P-T and the control group ($P = 0.016$; Bonferroni corrected) 5 weeks after surgery. No significant difference was observed between the control group and group P ($P = 0.102$) or between the control group and group P-T ($P = 0.335$) 3 weeks after surgery, and no significant difference was observed between group P and the control group ($P = 0.113$) or between group P and group P-T ($P = 0.582$) 5 weeks after surgery (Fig. 2).

**Degree of Adhesion between SRM and Sclera**

No significant difference was observed among three groups in terms of degree of adhesion between SRMs and sclerae 3

### Table 1. Numbers of Adjustable Eyes and Adjustment Tractional Forces and Distances

<table>
<thead>
<tr>
<th>Group</th>
<th>Time (wk)</th>
<th>Adjustment Possible (no. eyes)</th>
<th>Adjustment Not possible (no. eyes)</th>
<th>Force (g)</th>
<th>Length (mm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>C</td>
<td>3</td>
<td>0</td>
<td>5</td>
<td>*</td>
<td>*</td>
</tr>
<tr>
<td>P</td>
<td>3</td>
<td>5</td>
<td>5</td>
<td>54.00 ± 15.03</td>
<td>2.10 ± 0.55</td>
</tr>
<tr>
<td>PT</td>
<td>5</td>
<td>5</td>
<td>5</td>
<td>50.80 ± 21.94</td>
<td>2.35 ± 0.70</td>
</tr>
<tr>
<td></td>
<td>5</td>
<td>8</td>
<td>2</td>
<td>34.12 ± 9.92</td>
<td>2.31 ± 1.09</td>
</tr>
</tbody>
</table>

Values for force and length are mean ± SD. C, control; P, polytetrafluoroethylene; P-T, polytetrafluoroethylene/poly(lactide-co-glycolide) ± tranilast.

* Impossible to move the muscle because of adhesion.
weeks after surgery ($P = 0.099$). A significant difference was observed 5 weeks after surgery ($P = 0.033$), but the post hoc pairwise comparisons did not show any significant differences between three groups. No significant difference was observed between group P and the control group ($P = 0.505$) or between groups P and P-T ($P = 0.177$) 5 weeks after surgery. A considerable difference was observed between group P-T and the control group at this time but did not reach significance ($P = 0.026$; Bonferroni corrected; Fig. 2).

**Histologic Examination**

Histologic examination using with hematoxylin and eosin staining showed some degrees of inflammation with fibrosis around SRMs in the control group and in group P 3 weeks after surgery, and these were reduced 5 weeks after surgery. Mild inflammation and fibrosis were observed around SRMs in group P-T. Masson trichrome staining also revealed that fibrosis was least prominent in group P-T compared to that in group P or

**FIGURE 2.** Comparison of the degree of adhesion between groups. (A) Adhesion degree between superior rectus muscle and conjunctiva. The degree of adhesion was low in most eyes in group P-T at 3 or 5 weeks after surgery. A significant difference was observed between group P-T and group P ($P = 0.001$) 3 weeks after surgery and between group P-T and the control group ($P = 0.016$) 5 weeks after surgery. (B) Adhesion degree between superior rectus muscle and sclera. No significant difference was observed among three groups 3 weeks after surgery. At 5 weeks after surgery, the degree of adhesion was high in most eyes in group P and the control group. C, control; P, polytetrafluoroethylene; P-T, polytetrafluoroethylene/polylactide-co-glycolide + tranilast.

**FIGURE 3.** Light microscopic findings (Masson trichrome; original magnification, ×200) of the superior rectus muscle 3 weeks (A, C, E) and 5 weeks (B, D, F) after surgery. (A, B) Control group. (C, D) Group P. (E, F) Group P-T.
the control group (Fig. 3). No specific toxic reaction was observed in group P or group P-T.

**DISCUSSION**

PTFE has been used as pericardial and as abdominal patch graft material for several years and as an alloplastic xenograft for the treatment of eyelid and socket diseases or in glaucoma surgery.22–29 PTFE is chemically and biologically inert, nonantigenic, and resistant to infection, and it is easily cut, molded, and sutured.23 It is available as an expanded porous 1.0-mm-thick sheet or as a 0.1-mm nonporous thin surgical membrane. Nonporous 0.1-mm PTFE is soft and flexible and can be applied to complex shapes without difficulty. Moreover, because of its thinness, it is well tolerated by patients and generally does not cause discomfort. Given these advantages, we considered PTFE the most suitable nonabsorbable material choice for delaying adjustment.

The ideal drug-release device for our purpose would fulfill the following criteria.27 First, it should provide a sustained and relatively uniform therapeutic drug release in a reliable and predictable manner over a required period. Second, the device should be easily implantable or injectable but remain stable and not migrate within the eye. Third, the device should have a long shelf life, and it should be easily handled and sterilize. Moreover, because the conjunctival incision must be reopened to adjust sutures, unlike other intraocular devices, it does not have to be biodegradable. None of the sustained delivery systems introduced to date fulfill these criteria.

In the present study, a 0.1-mm-thick PTFE film coated with PLGA was used. PLGA film is difficult to manipulate because it tends to roll up and stick to itself,11 but PTFE coated with PLGA is easier to handle. Our previous study suggested that PTFE alone might allow adjustment to be delayed for up to 4 weeks after surgery in 40% of rabbit eyes26 and that the combined use of PTFE and 5-fluorouracil, or the addition of viscoelastic solution (Viscoat; Alcon Laboratories, Fort Worth, TX) could allow adjustment to be delayed for up to 4 weeks after surgery in 80% of rabbit eyes.13

In this study, we evaluated adjustability 3 weeks and 5 weeks after surgery based on a release profile of tranilast, which showed sustained release up to 4 weeks. Only group P-T was found to be significantly more adjustable than the control group 3 and 5 weeks after surgery. Moreover, only group P-T showed significantly less adhesion between SRMs and conjunctivae 3 and 5 weeks after surgery. In addition, the present study produced favorable results without the need for frequent topical steroid instillation. These results demonstrate that PTFE/PLGA, which allowed sustained tranilast release, effectively prevented adhesion development and improved adjustability after strabismus surgery in our rabbit model.

Tranilast is known to inhibit the release of histamine, leukotriene, interleukin, and TGF-β and to suppress the initial stages of the wound-healing processes such as vascular leakage, fibrin deposition, and migration of monocytes and fibroblasts.28–30 In this study, the histopathologic findings of group P-T showed minimal inflammation and fibrosis 3 and 5 weeks after surgery even though we did not evaluate the degree of inflammation or fibrosis quantitatively compared with other groups.

Tranilast has also been evaluated in some ocular diseases. Chikara et al.31 reported the use of topical tranilast after filtering surgery reduced intraocular pressure and increased the size of the bleb. In addition, an in vitro study showed tranilast inhibited rabbit tenon capsule fibroblast and collagen synthesis.32 Tranilast is also thought to reduce corneal haze after photorefractive keratectomy.33

**References**

18. Lee JP, Choung HK, Kim CK, Hwang JM. Polytetrafluoroethylene/poly lactide-co-glycolde laminate containing dexamethasone al-


