Uveal melanoma arises from neural crest-derived melanocytes of the uveal tract and is the most common primary intraocular malignant tumor in adults with an incidence of four to seven individuals per 1 million/y in the United States. Clinical presentation varies depending on the size and location of the tumor. Median age at presentation is 55 years of age, and the majority of patients are Caucasians. Metastasis develops in up to 50% of primary uveal melanoma patients, usually through hematogenous spread. Regional lymphatic dissemination occurs rarely, due to the relative lack of lymphatic drainage of the choroid. The most common site of metastasis is the liver (occurring in as many as 90% of patients with metastatic uveal melanoma), and the median survival of those patients is approximately 4 to 5 months. Approximately 50% of patients with liver metastasis also have extrahepatic involvement, the most common sites being lung (30%), bone (23%), and skin (17%). Factors predicting metastatic disease are large tumor diameter, ciliary body involvement, extrascleral extension, epithelioid melanoma histology, vascular matrix pattern (such as closed loops), high mitotic rate, microvascular density, monosomy 3, and class 2 gene expression profile.

While radical treatment of uveal melanoma consists of enucleation, the most common treatments are conservative, such as brachytherapy and external irradiation (e.g., proton beam). Survival rates and risk of metastasis are similar with either enucleation or radiation. Despite good local control of uveal melanoma, the treatment of metastatic disease is still limited due to its resistance to conventional systemic chemotherapy. Many drugs,
such as imatinib, bevacizumab, and trametinib (a reversible, selective allosteric inhibitor of MEK1 and MEK2)\(^5\) are presently under investigation along with intrahepatic injection or surgical intervention.\(^3\) However, there is insufficient evidence that any pharmacologic treatment prolongs survival in patients with metastatic uveal melanoma.\(^19\)

Adenosine monophosphate–activated protein kinase (AMPK) is a heterotrimeric serine/threonine protein kinase that is a major sensor and regulator of cellular and whole-body energy levels and stress.\(^{20−24}\) Its activity is regulated by conditions that deplete cellular ATP and elevate AMP levels (such as hypoxia, exercise, ischemia, glucose deprivation, and heat shock),\(^25\) and also by some hormones such as leptin,\(^26\) adiponectin,\(^27\) catecholamine,\(^28\) and IL-6.\(^29\) Adenosine monophosphate–activated protein kinase upstream protein kinase liver kinase B1 (LKB1)\(^30,31\) is a tumor suppressor that is mutated in Peutz-Jegher syndrome. Its downstream effectors also involve the tumor suppressor tuberous sclerosis complex (TSC2) and the mammalian target of rapamycin (mTOR), which are known to be important factors in cell-cycle progression and tumor formation.\(^32,33\) Although tuberous sclerosis complex (TSC2) and the mammalian target downstream effectors also involve the tumor suppressor a tumor suppressor that is mutated in Peutz-Jegher syndrome. Its activation,\(^39\) and ischemia/reperfusion injury.\(40\) Aminoimidazole carboxamide ribonucleotide–mediated activation of AMPK has been shown to inhibit proliferation and induce apoptosis in many pharmacologic activators of AMPK exist, 5-aminoimidazole-4-carboxamide riboside (aminoimidazole carboxamide ribonucleotide [AICAR]) was the first compound reported to activate AMPK both in intact cells and in vivo.\(^34,35\) Aminoimidazole carboxamide ribonucleotide is taken into cells by adenosine transporters and then converted by adenosine kinase to the monophosphorylated form, 5-aminimidazole-4-carboxamide-1-D-ribofuranosyl-5′-monophosphate (ZMP), which mimics an increase of AMP intracellular levels. In addition to its AMPK-dependent effects, AICAR can also be converted to inosine, which acts in an AMPK-independent manner to increase cellular adenosine concentration.\(^34−36\) The toxicity of AICAR is low or not apparent when given in intraperitoneal doses up to 500 mg/kg/day for 4 weeks in mice.\(^34\)

Adenosine monophosphate–activated protein kinase activation has been reported to have both prosurvival and proapoptotic effects depending on the environment and the stimulus; for example, AMPK activation has been shown to be antiapoptotic in situations of hyperglycemia,\(^38\) glucose deprivation,\(^39\) and ischemia/reperfusion injury.\(40\) Aminoimidazole carboxamide ribonucleotide–mediated activation of AMPK has been shown to inhibit proliferation and induce apoptosis in retinoblastoma cells (both in vitro and in vivo),\(^31,42\) neuroblastosoma cells,\(^43\) childhood acute lymphoblastic leukemia cells,\(^44\) glioblastoma cells,\(^45\) multiple myeloma cells,\(^46\) prostate cancer cells, breast cancer cells,\(^47\) hepatic cancer cells,\(^48\) and colon cancer cells.\(^48\) Various mechanisms have been demonstrated, such as upregulation of p53,\(^44\) increased expression of cell-cycle inhibitory proteins p21 and p27,\(^44\) inhibition of the Akt/mTOR/P70S6K pathway,\(^44\) decrease of cyclins A and E,\(^43\) inhibition of nuclear factor kappa-B (NF-kB) activity,\(^36,48\) and decreased angiogenesis.\(^12\) The various effects of AMPK on survival or growth inhibition likely depend on cell type, duration of AMPK activation, cellular events following external stimuli, and/or downstream regulated pathways of AMPK.

In the present study, we investigated the effects of AICAR on cell proliferation and its mechanism of action in vitro in three uveal melanoma cell lines.

Materials and Methods

Chemicals

Aminoimidazole carboxamide ribonucleotide was purchased from Toronto Research Chemicals (Toronto, Ontario, Canada). Dipyridamole and 5-iodotubercidin (iodo) were purchased from Sigma-Aldrich (St. Louis, MO, USA). Aminoimidazole carboxamide ribonucleotide was dissolved in RPMI 1640 medium (Invitrogen [Life Technologies], Carlsbad, CA, USA) at a concentration of 40 mM (stock solution) and stored at −20°C until used. Dipyridamole and iodo were prepared fresh from stock solutions and diluted with growth medium. 3(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) was purchased from Sigma-Aldrich. The following primary antibodies were purchased from Cell Signaling Technology (Danvers, MA, USA): phospho-ACC (Ser-79), phospho-S6 ribosomal protein (Ser-235/236), CDK2, CDK4 (DCS156), p21 Waf1/Cip1 (12D1), p27Kip1 (D37H1), p53, LC3B, phospho-4E-BP1 (Ser-65), PCNA (PC10), phospho-p44/42 MAPK (Erk1/2) (Thr202/Tyr204) (D13.14.4E), and β-actin (1:5E5).

Cell Culture

The primary cell lines OCM 3 (aka SkMel28),\(^49\) 92.1.50 MEL 270,\(^31\) and MEL 202\(^52\) were grown in RPMI medium supplemented with 10% fetal bovine serum (FBS; Invitrogen), penicillin (100 μg/mL; Invitrogen), and streptomycin (100 μg/mL; Invitrogen). MEL 270 and MEL 202 were additionally supplemented with 1% minimal essential medium (MEM) vitamin solution and 1% MEM nonessential amino acids (Invitrogen). Cells were incubated at 37°C in a humidified atmosphere containing 5% CO2 and split when they reached approximately 90% to 95% confluence.

Measurement of Cell Growth by MTT Assay

Cell viability was assessed by MTT assay. Cells were cultured in 96-well plates at a density of 4000 cells/well in 150 μL growth medium and were incubated with AICAR (1, 2, and 4 mM), dipyridamole (2 μM), or 5-iodotubercidin (0.1 μM) at 37°C for 3 and 5 days. On days 1, 3, and 5 of treatment, the culture medium was aspirated and 60 μL MTT (5 mg/mL in RPMI 1640 without phenol red; Sigma-Aldrich) was added to each well. After incubation for 2 hours at 37°C, 130 μL of dimethyl sulfoxide was added. The absorbance or optical density (OD) at 595 nm was measured using a microplate reader (Molecular Devices, Sunnyvale, CA, USA). For each treatment, cell growth was evaluated as a percentage using the following equation:

\[
\frac{(1)}{(1)} = \left(\frac{OD_{95\%} - OD_{95\%}}{OD_{95\%} - OD_{95\%}}\right) 
\]

Cell Cycle Assessment by Flow Cytometry

Cellular DNA content was assessed by flow cytometry. Cells were cultured in 10-cm plates at a density of 2.5 million cells/plate in 10 mL growth medium, and were treated with 1 and 2 mM AICAR for 1, 3, and 5 days. After drug treatment, the cells were trypsinized, spun at 200g for 5 minutes, and washed twice with 1-mL cold PBS. While the cells were continuously vortexed, 2 mL ice-cold 75% ethanol was added slowly, and the cells were then fixed overnight. On the day of measurement, cells were spun, resuspended in 2 mL PBS with the addition of 100 μL of DNase-free RNase A (200 μg/mL; Invitrogen), and incubated at 37°C for 30 minutes. Then, 100 μL of 1 mg/mL propidium iodide (Invitrogen) was added, and the cells were incubated at room temperature for 10 minutes. The samples were stained on a Becton Dickinson FACScan (Becton Dickinson, Franklin Lakes, NJ, USA). The sub-G1 peak was quantified and represented the nonviable cell population.

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Western Blot Analysis

After 24 hours of incubation in the presence or absence of AICAR, medium was aspirated and the plate was washed three times with cold PBS and kept in −80°C overnight. On the next day, 500 µL of 1× lysis buffer (Cell Signaling Technology) containing protease and phosphatase inhibitor cocktail (Roche, Indianapolis, IN, USA) were added per 10-cm dish, incubated for 5 minutes on ice, and cells were scraped. Extract was centrifuged for 10 minutes at 14,000 g in a cold microcentrifuge. The supernatant was removed, and lithium dodecyl sulfate sample buffer (Invitrogen) containing dithiothreitol (American Bioanalytical, Natick, MA, USA) was added to equal amounts of total protein from each sample and heated at 90°C for 5 minutes. Samples were loaded onto a NuPAGE 4–12% Bis-Tris Gel (Invitrogen) and then transferred to a polyvinylidene fluoride (PVDF) membrane (0.45 µm; Millipore, Billerica, MA, USA). The membranes were incubated overnight with primary antibody at 4°C with gentle shaking. Primary antibodies were diluted 1:1000 in 5% wt/vol BSA, Tween-20 (TBST) with exception of the antibodies for p53, CDK4 and PCNA, which were diluted in 5% nonfat dry milk, TBST. The blotted membranes were washed three times (5 minutes/wash) with TBST and incubated for 45 minutes at room temperature with horse radish peroxidase-labeled anti-rabbit or anti-mouse secondary antibody (1:100,000; Jackson ImmunoResearch, West Grove, PA, USA). The membranes were washed three times (5 minutes/wash) in TBST, and immune reactive bands were visualized by enhanced chemiluminescence (ECL) and exposure onto Fuji RX film (Fujifilm, Tokyo, Japan) for approximately 5 minutes.

Quantitative Real-Time RT-PCR

After 24 hours of incubation in the presence or absence of AICAR, the medium was aspirated and plates were washed with cold PBS. Cellular RNA was extracted and purified with the RNeasy Micro kit (Qiagen, Valencia, CA, USA). Ribonucleic acid was further cleaned with an additional DNase I digestion step according to the manufacturer’s instructions. Reverse transcription was performed for equal RNA amounts (4 µg, as measured by ultraviolet spectrophotometry) with oligo dT primer (Invitrogen) and Superscript II (Invitrogen). Complementary DNA (100 ng) was used for each of the three replicates for quantitative PCR. Human cyclin A1, cyclin A2, cyclin D1, cyclin D3, cyclin E1, cyclin E2, and 18S, and β-actin (as endogenous controls) were amplified with commercially designed Taqman gene expression assays (Applied Biosystems, Foster City, CA, USA) and the Taqman universal PCR master mix (Applied Biosystems). Quantitative expression data were acquired and analyzed with a Step One Plus real-time PCR system (Applied Biosystems).

Statistical Analysis

The results are expressed as the mean ± SEM. Data were analyzed by Student’s t-test or ANOVA of the repeated experiments with Prism software (GraphPad Software, San Diego, CA, USA). For all analyses, significance was assigned at P < 0.05.

RESULTS

AICAR Inhibits the Growth of Uveal Melanoma Cells

To study the effect of AICAR on the growth and metabolism of uveal melanoma cells, one skin melanoma cell line (OCM 3) and three uveal melanoma cell lines (92.1, MEL 270, and MEL 202) were treated with AICAR (1, 2, and 4 mM) for 3 and 5 days. Their metabolism and growth was evaluated using the MTT assay. Aminoimidazole carboxamide ribonucleotide inhibited their growth in a time- and dose-dependent manner (P < 0.05 for all cell lines; Fig. 1, Supplementary Fig. S1).

Cellular uptake of AICAR occurs via adenosine transporters. To confirm that the inhibition of uveal melanoma cells was dependent on receptor-mediated uptake of AICAR, we pretreated cells with dipyridamole, which blocks adenosine transporters and prevents uptake of AICAR into the cells. As a negative control, dipyridamole treatment alone did not affect cell metabolism and growth. In contrast, treatment of uveal melanoma cells with dipyridamole plus AICAR abolished the inhibitory effect of AICAR in all cell lines (P < 0.05), indicating that surface adenosine receptors are expressed on uveal melanoma cells and mediate the uptake and effects of AICAR (Fig. 2A, Supplementary Fig. S2A).

Antiproliferative Effects of AICAR are Mediated at Least Partially via the AMPK Pathway

Since AICAR has been reported to be able to inhibit cell growth and proliferation via an AMPK-independent mechanism, it is...
important to determine whether AMPK activation coincides with the antiproliferative effects of AICAR on uveal melanoma cells. To confirm that AICAR treatment of uveal melanoma cells was associated with AMPK activation, we examined the phosphorylation of acetyl-CoA carboxylase (ACC), the downstream target of AMPK. Cells treated with AICAR (1 and 2 mM) showed an increase of phosphorylated ACC (Fig. 3A, Supplementary Fig. S3A). To confirm that ACC phosphorylation was due to intracellular AICAR, cells were pretreated with dipyridamole before AICAR. Blocking adenosine receptors and AICAR entry into the cells with dipyridamole inhibited ACC phosphorylation (Fig. 3B, Supplementary Fig. S3B). These data indicate that the AICAR-mediated inhibition of uveal melanoma cells coincides with activation of the AMPK pathway.

Other investigators have reported that once AICAR enters the cells it can be converted to either inosine or ZMP. Inosine can inhibit cells via an AMPK-independent pathway, whereas ZMP activates the AMPK pathway. Aminoimidazole carboxamide ribonucleotide is converted to ZMP by adenosine kinase, but this conversion is blocked by iodo. To determine whether uveal melanoma cells inhibition by AICAR coincides with the conversion of AICAR to ZMP, we pretreated the cells with iodo prior to AICAR administration. Activation of AMPK was assessed by examination of ACC phosphorylation. Although activation of AMPK was shown to be effectively blocked by iodo treatment as judged by phosphorylated ACC immunobLOTS (phosphorylated ACC ± iodo; inhibition at \( P < 0.05 \); Fig. 3C, Supplementary Fig. S3C), a significant, but not complete reversal of AICAR-mediated uveal melanoma cell growth inhibition was observed in OCM 3, 92.1, and MEL 270 cell lines, but not MEL 202 (Fig. 2B, Supplementary Fig. S2B), indicating that AMPK activation by ZMP is only partially responsible for the observed inhibitory effects of intracellular AICAR.

### AICAR Causes Cell Cycle Arrest in S Phase of Uveal Melanoma Cell Lines

The reported effects of AICAR on the cell cycle have been variable depending on the cell type studied. To examine the effect of AICAR on uveal melanoma cell cycle profiles, cells were treated with AICAR (1 and 2 mM) for 1, 3, and 5 days, and the cell-cycle phase was analyzed for nuclear DNA content by propidium iodide staining and flow cytometry. Compared with control cells, AICAR treatment resulted in accumulation of cells in S phase (Fig. 4, Supplementary Fig. S4) in a dose-dependent manner.

### AICAR Decreases the Levels of Cyclins A and D in Uveal Melanoma Cells

Cell cycle progression is controlled by specific cyclins. Given the effects shown previously of AICAR on uveal melanoma cell cycle regulation, we wanted to check whether that effect was mediated by changes in the levels of the appropriate cyclins. After the cells were treated with AICAR (1 and 2 mM) for 24 hours, quantitative RT-PCR analysis showed a significant dose-dependent decrease of cyclins A1 and D1 in all cell lines; in addition to cyclin D3 in MEL 270 and cyclins A2 and E2 in MEL 202 (Fig. 5, Supplementary Fig. S5). These results suggest that in uveal melanoma cells, AICAR-induced S phase arrest might be associated with decreasing levels of cyclin proteins.
AICAR Does Not Affect the Levels of the Cyclin-Dependent Kinases CDK2 and CDK4, CDK Inhibitor p27, p21, Tumor Suppressor Protein P53, PCNA, and MAPK Pathway

Other cell cycle progression regulators have been reported to be affected by AICAR in various cell types. We wanted to check whether AICAR affects some of these regulators in uveal melanoma cells. We thus examined its effect on CDK2, CDK4, CDK inhibitor p27, p21, tumor suppressor protein p53, and PCNA. As shown in Figure 6 and Supplementary Figure S6, AICAR had little or no effect on the expression of the mentioned cell cycle regulators except the significant increase in p53 levels in MEL 270 cell line. In addition, we did not see change in the MAPK pathway, which has been reported to play a role in the pathogenesis of uveal melanoma.

AICAR Downregulates 4E-BP1 Phosphorylation but Not S6 Kinase or the Macroautophagy Marker LC3B in Uveal Melanoma Cells

The mTOR pathway has been demonstrated to be one of the major pathways controlling cell proliferation and autophagy. Adenosine monophosphate–dependent kinase directly and indirectly inhibits mTOR/Raptor,60 directly phosphorylates Ulk1, and promotes autophagy.61–63 The nonselective type of autophagy called macroautophagy is thought to be regulated and inhibited by S6 kinase, a downstream effector of mTOR.64–66

Aminoimidazole carboxamide ribonucleotide’s effects on multiple cell types have been shown to be mediated through the mTOR pathway and autophagy.67–70 In contrast to our prior work on human retinoblastoma cells,41,42 Aminoimidazole carboxamide ribonucleotide did not inhibit the phosphorylation of ribosomal protein S6, a downstream effector and a
measure of mTOR activity (Fig. 6, Supplementary Fig. S6). However, AICAR downregulated 4E-BP1 phosphorylation (another marker of mTOR activity) in OCM 3, 92.1, and MEL 270 cell lines, but not in MEL 202 ($P < 0.05$; Fig. 7, Supplementary Fig. S7). In addition, the macroautophagy marker LC3B was found to be significantly increased only in OCM 3 cell line (Fig. 6, Supplementary Fig. S7). This suggests that the AICAR’s effects in uveal melanoma on the mTOR pathway and autophagy are more complex than in other cell lines.

**DISCUSSION**

In this study, we demonstrated that AICAR, a pharmacologic activator of AMPK, can induce S phase cell-cycle arrest and inhibit growth in three human uveal melanoma cell lines. Dipyridamole, an adenosine transporter inhibitor, abolished these AICAR-mediated effects by preventing its cellular uptake. The adenosine kinase inhibitor iodotubericidin, which inhibits the enzyme responsible for converting AICAR to ZMP, abated AMPK activation (demonstrated by ACC phosphorylation) and blocked AICAR’s growth inhibitory effects, suggesting that these effects are mediated by intrinsic mechanisms and at least partially by AMPK activation.

Previous reports from us and other laboratories indicate that the cell type determines the AICAR effects on cell cycle. Aminoimidazole carboxamide ribonucleotide’s treatment of various cancer cell lines has showed arrest either in the S phase, $36,46$ G1 phase, $57$ and/or an increase in the sub-G0/G1 population. $41,48$ An increase in the S-phase population was observed upon treating three uveal melanoma cell lines with AICAR, which also caused downregulation of cyclins A1 and D1. This is consistent with S phase arrest, as cyclins A1 and D1 control progression through S phase. We also observed downregulation of other cyclins in MEL 270 and MEL 202 cell lines.

The mechanisms of AICAR’s inhibitory effects vary depending on the cell line being studied, and multiple mechanisms have been shown to play a role in the inhibiting effects of AICAR. Adenosine monophosphate–dependent kinase activity was upregulated and/or necessary in retinoblastoma, multiple...
myeloma, colon, breast, prostate, and hepatic cancer cell lines, whereas AMPK activity was nonessential in studies of Jurkat cells, myelogenous leukemia, and neuroblastoma cell lines. Our results indicated that, in uveal melanoma cells, AMPK activity was at least partially required for the inhibitory effects of AICAR in two uveal melanoma cell lines (92.1 and MEL 270) and the skin melanoma cell line OCM 3, but not in MEL 202 cell line.

Further investigation of the growth inhibitory mechanisms of AICAR revealed more differences depending on the cell lines studied. Aminoimidazole carboxamide ribonucleotide treatment has been shown to inhibit glioblastoma cells by inhibiting lipogenesis, triggered apoptosis by inhibiting NF-κB pathway in colon cancer cells, and inhibited proliferation by upregulating the cell cycle inhibitor protein p21 in C6 glioma cells and acute lymphoblastic leukemia cells; however, it inhibited p21 in retinoblastoma cells. Aminoimidazole carboxamide ribonucleotide has been shown to increase p27 and decrease PCNA in C6 glioma cells. In contrast to these studies, the effects we observed in AICAR-treated uveal melanoma cells did not occur through any of these mechanisms, except for the increase in p53 in MEL 270.

Aminoimidazole carboxamide ribonucleotide has been shown to be an exercise mimetic and demonstrates antiinflammatory properties, in addition to prosurvival effects in normal cells under stress. The mechanisms responsible for these effects are not fully understood, but they likely involve activation of AMPK. It is possible that the various effects of AICAR and AMPK depend on the specific cell type, cellular events following external stimuli, duration of AMPK activation, and/or downstream-regulated pathways of AMPK.

Research on the antitumor effects of AICAR-induced AMPK activation is becoming an important area of investigation because of its link with tumor suppressors. The tumor suppressor LKB1 is an upstream activator of AMPK, and the encoding gene is mutated in Peutz-Jeghers syndrome, an autosomal dominant disease characterized by hamartomatous polyp growth and predisposition to cancers of the gastrointestinal tract. Tuberous sclerosis 2 (TSC2) is known to be downstream of activated AMPK; it forms a complex with TSC1 and inhibits mTOR, leading to negative regulation of cell growth. Mutations of the TSC2 gene are associated with tuberous sclerosis, which in humans is associated with...

**Figure 5.** Aminoimidazole carboxamide ribonucleotide decreases the levels of different cyclins in uveal melanoma cells. 92.1 (A), MEL 270 (B), and MEL 202 (C) uveal melanoma cells were treated with AICAR at a concentration of either 1 or 2 mM for 24 hours. Quantitative RT-PCR analysis showed decrease of cyclins A1 and D1 in all cell lines, cyclin D3 in MEL 270 and cyclins A2 and E2 in MEL 202-treated cells in comparison with control cells. Significance (*) is assigned at $P < 0.05$. 

...
hamartomas and an increased risk of cancers. Mammalian target of rapamycin phosphorylates 4E-BP1, leading to release of eIF4E and allowing initiation of translation. Hyperphosphorylation of 4E-BP1 has been reported to be a marker of poor prognosis and a potential target for the treatment of cutaneous melanoma, breast cancer, and astrocytoma.72–74 We observed decreased phosphorylation of 4E-binding protein 1 (4E-BP1), a downstream pathway of mTOR, in three of the four cell lines tested. However, S6 kinase, another downstream effector of mTOR, was not downregulated after AICAR treatment in contrast to our previous study in retinoblastoma41,42 and the study by Rattan et al.36 in C6 glioma cells, suggesting that AICAR’s effects in uveal melanoma on the mTOR pathway may be more complex than in other cell lines.

Adenosine monophosphate–dependent kinase activation has been reported to induce autophagy by suppressing mTOR pathway, and thus suppressing the macroautophagy inhibitor S6 kinase, and by directly phosphorylating proautophagy protein Ulk1.60,64–66 The role of autophagy in cancer is still debated, and can be either detrimental or protective.75 Adenosine monophosphate–dependent kinase induction of autophagy has been thought to contribute to cell death in colorectal HT-29 cells,76 and AICAR has been shown to induce cell death and autophagy stimulation in chronic myelogenous leukemia cell lines.70 We failed to observe any significant and consistent effects of AICAR on the autophagy marker LC3B; thus, the possibility remains that other mechanisms are responsible for the inhibition of uveal melanoma cells.

Although advances in therapy for uveal melanoma have led to significant success in local control, metastasis remains a significant problem with a lack of effective therapies. This underscores the need for the development of new targets and less toxic therapies.

In summary, our results show that AICAR, after entering the cells, inhibits uveal melanoma cell growth at least partially through activation of AMPK, inhibition of 4E-BP1 phosphorylation, and downregulation of cyclins A1 and D1. Moreover, other studies have shown that AICAR, when administered in nonchronic situations, has low toxicity, displays antiinflammatory properties, and acts as an exercise mimetic.37 In addition AICAR (also known as acadesine) is already in human clinical trials for B Cell leukemia and early phase I/II study results have shown trends of efficacy; reduction of peripheral chronic lymphocytic leukemia (CLL) cells and reduction in lymphadenopathy were observed with blood levels close to 1 mM.77 Together, these data indicate that AICAR has potential as a novel targeted therapy with low toxicity for uveal melanoma.
**FIGURE 7.** Antiproliferative effect of AICAR on uveal melanoma cells is mediated via inhibition of 4E-BP1 phosphorylation in 92.1 and Mel 270, but not in Mel 202 cells. Western blot analysis of P-4E-BP1 in 92.1, Mel 720, and Mel 202 cells treated with AICAR at a concentration of either 1 or 2 mM for 24 hours. Density values of the bands are graphically expressed relative to control. Multiple bands represent separate biological samples. Significance (*) is assigned at $P < 0.05$.

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