Effect of Prostaglandin F2α on Aqueous Humor Dynamics of Rabbit, Cat, and Monkey

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Topical administration of prostaglandin F2α (PGF2α) produced a reduction in intraocular pressure in eyes of rabbits, cats, and cynomolgus monkeys. In rabbit eyes at 5 or 6 hr, 50 µg, 100 µg, or 250 µg of PGF2α caused a significant intraocular pressure reduction with a small miotic effect. Treatment with 500 µg, 750 µg, or 1000 µg of PGF2α lowered intraocular pressure significantly in cat eyes for at least 24 hr with the development of profound pupillary constriction. Administration of 500 µg, 750 µg, or 1000 µg of PGF2α produced a significant reduction of intraocular pressure in monkey eyes lasting at least 24 hr, with an initial hypertensive phase and a small decrease in pupillary diameter in the treated eyes. Tonography revealed an increased facility of outflow simultaneous with the reduction of intraocular pressure in the eyes of cats and monkeys. These increases of outflow facility could not explain completely the reductions in intraocular pressure. The aqueous humor flow measured by fluorophotometry was unaltered in both species, and possible reasons for this finding are discussed. Anterior chamber aqueous humor protein was significantly higher in cat eyes topically treated with 750 µg of PGF2α than in the diluent-treated fellow eyes. Invest Ophthalmol Vis Sci 25:1087-1093, 1984

Early studies of the effect of prostaglandins (PGs) on intraocular pressure led to the general conclusion that PGs, administered topically or systemically, elevated intraocular pressure in rabbits, cats, and monkeys.1-5 More recently, some studies have shown that topical application of either PGE2 or PGF2α effectively reduced the intraocular pressure in rabbits, cats, and monkeys.6-8 Those studies suggested that PGs, especially PGF2α, and/or its analogues, may provide a new therapeutic approach to the clinical control of intraocular pressure and the treatment of glaucoma.

The present study was designed to investigate further the mechanism of the hypotensive effect of PGF2α on rabbit, cat, and monkey eyes.

Materials and Methods

Adult, albino, unanesthetized rabbits, 2–3 kg, were restrained. Eleven adult cats, 2.5–3.5 kg, and eight, adult, cynomolgus monkeys, 4–5 kg, were lightly tranquilized with 5–10 mg/kg of ketamine. The cats were restrained, and the monkeys were kept in primate chairs throughout each experiment.

Intraocular pressure was measured under 0.5% topical proparacaine hydrochloride anesthesia using a manometrically calibrated Alcon pneumatonometer. New animals were acclimated to the tonometer by undergoing several readings the day before they were to be used in an experiment. Two sets of baseline readings were taken each day before 9 AM.

Pupillary diameters were measured with a millimeter ruler in normal room light. In cats, the horizontal (shorter) diameter always was recorded.

The aqueous flare and cellular response in the anterior chamber were assessed by slit-lamp examination and rated from 0 to 3 (aqueous flare: 0 = no Tyndall effect; 1+ = slight Tyndall effect; 2+ = moderate to dense Tyndall effect; 3+ = dense Tyndall effect with fibrin clots; cellular response: 0 = no cells apparent; 1+ = few cells; 2+ = many cells; 3+ = cell clumps).

Following these baseline observations, a 5 mg per ml solution of PGF2α (each ml of this solution contains prostaglandin F2α, tromethamine salt equivalent to 5 mg prostaglandin F2α, and benzyl alcohol, 9.45 mg, added as a preservative. The Upjohn Co. (Kalamazoo, MI), diluted with normal saline to various concentrations, was applied topically to one eye of each animal. As topical application of an aqueous solution containing 9.45 mg per ml of benzyl alcohol did not alter the intraocular pressure in our trials with cynomolgus monkeys, we used an equal volume of normal saline applied to the contralateral eye as the control. All the drugs were made up just prior to their administration. The following amounts of PGF2α were applied: rabbits—1 µg in 1 µl, 5 µg in 1 µl, 25 µg in 5 µl, 50 µg...
in 50 μl, 100 μg in 50 μl, and 250 μg in 50 μl; cats and monkeys—250 μg, 500 μg, 750 μg, 1000 μg as 250 μg in 50 μl given one, two, three, or four times, respectively, 3–5 min apart. Repeat, intraocular pressure measurements were made at 0.5, 1, 1.5, 2, 3, 4, 5, 6, and 24 hr after instillation of PGF₂α.

Tonography was performed with an electronic tonometer (Alcon EDT-103) in 21 cats and 20 monkeys. Baseline outflow facility was determined at 8:30 AM–9 AM. PGF₂α (750 μg in cats and 500 μg in monkeys) was applied randomly to one eye and an equal volume of normal saline to the contralateral eye 2 hr after baseline measurements. The tonography results were obtained at 2 hr (cats) or 4 hr (monkeys) after instillation of PGF₂α. Tonography values were approximated from the 1955 Friedenwald tables.

Aqueous humor flow was estimated using a fluorophotometric technique on 14 cats and 10 monkeys. The fluorescein iontophoresis was done at 4 PM and fluorescence measurements were made from 9 AM–2 PM on the following day. The iontophoresis was carried out in the central 4 mm of the cornea with an electrode of 10% fluorescein in 2% agar. A current of 200 μA was used for 5 min. Fluorophotometric measurements of the cornea and anterior chamber were repeated at about 60-min intervals. Five to six such measurements were made. Following these baseline measurements, on another day, PGF₂α (750 μg in cats and 500 μg in monkeys) was topically applied to one eye of each animal at about 8:30 AM. An equal volume of normal saline was applied to the control eye. The iontophoresis was carried out at 4 PM on the preceding day as described above. Fluorophotometric measurements were taken from 1–6 hr after instillation of PGF₂α. The cornea and anterior chamber readings were divided by the reference filter reading and the ratio (F) was recorded. For each animal, the natural logarithm of F was plotted versus time. The lines of best fit and their slopes were calculated by the least-squares method.

The value of aqueous flow was calculated by the mathematical assumptions of Yablonski and co-workers. The value of A used for each eye was midway between the absolute values of the slopes of the anterior chamber and cornea lines of best fit. The value of Fe/Fa was determined from the corresponding lines of best fit at 2 hr (cats) or 4 hr (monkeys) after PGF₂α administration. Values of 853 μl for anterior chamber volume and 296 μl for cornea volume were used in the calculations. Values of 106 μl for anterior chamber volume and 50 μl (unpublished data, M. E. Yablonski and J. B. Serle) for cornea volume in monkeys were used in the calculations.

Seven hundred fifty micrograms of PGF₂α were instilled in one eye of awake, restrained cats, control solution in the other eye. Two hours later, under ketamine anesthesia, a 25-gauge needle was inserted through clear cornea and aqueous humor withdrawn. Care was taken to avoid the iris and lens. Aqueous humor protein concentrations were measured by the method of Lowry and co-workers.

These experiments adhered to the ARVO resolution on the use of experimental animals in research.

**Results**

**Intraocular Pressure**

*Rabbits.* PGF₂α administered topically to rabbit eyes often induced a biphasic intraocular pressure response:
a relatively short, initial, hypertensive phase followed by prolonged hypotony. Dose–response relationships could be demonstrated (Fig. 1). Topical application of 100 µg or 250 µg of PGF₂α produced a significant (P < 0.01) initial increase in intraocular pressure. Topical application of all doses of PGF₂α produced a significant (P < 0.05) ocular hypotony at 5 or 6 hr. The greatest hypotensive response was observed in eyes given 250 µg of PGF₂α.

**Cats.** Topical application of 500–1000 µg PGF₂α to the eyes of cats produced a significant (P < 0.05) decrease in intraocular pressure, as compared with the pressure of the control eyes, occurring between 30 min–24 hr after PGF₂α administration. The greatest hypotensive response was observed in eyes given 750 µg of PGF₂α at 2 hr (P < 0.001). There was no transient ocular hypertensive response in cats. Dose–response relationships could be shown (Fig. 2).

**Monkeys.** Topical application of 250 µg, 500 µg, 750 µg, or 1000 µg of PGF₂α to one eye of monkeys resulted in a biphasic intraocular pressure response: a relatively short initial hypertensive phase followed by a prolonged hypotony (Fig. 3). The maximum rise of the pressure occurred at 30 min. The intraocular pressure then rapidly decreased. The maximum ocular hypotensive response occurred after topical application of 500 µg of PGF₂α, with a significant (P < 0.001) decrease in intraocular pressure of 4 mmHg at 4 hr, as compared with the pressure of the control eyes. The intraocular pressure was significantly (P < 0.05) reduced up to 24 hr by 500–1000 µg PGF₂α.

**Miotic Response**

**Rabbits.** Topical application of 50 µg or 100 µg of PGF₂α produced a miotic response (P < 0.05) of 1 mm at 1.5 hr, which returned to baseline values at 5 hr (Fig. 4).

**Cats.** Topical administration of 500 µg, 750 µg, or 1000 µg of PGF₂α caused significant (P < 0.01) miotic responses similar in magnitude (Fig. 5). A dose of 500 µg of PGF₂α produced an apparently maximum miotic response (9 mm decrease in pupillary diameter) at 1 hr, the pupillary diameter 7 mm less than the control...
Fig. 4. Effects of topically applied PGF$_{2\alpha}$ on the pupillary diameter of rabbit eyes as compared with the pupillary diameter of control eyes. The greatest SE was ±0.4 mm.

Eyes for 5 hr, followed by redilation to near baseline values at 24 hr.

Monkeys. Topical application of PGF$_{2\alpha}$ doses produced a small decrease in pupillary diameter in the treated eyes and an increase in the control eyes. The effects of topically applied PGF$_{2\alpha}$ on the pupillary size of monkeys occurred between 15 min and 4 hr after PGF$_{2\alpha}$ administration (Fig. 6). The miotic response of the treated eyes and the dilation of the pupil of the control eyes were significant ($P < 0.02$) as compared with the baseline values 0.5 hr after the application of 1000 /ig of PGF$_{2\alpha}$. Topical application of PGF$_{2\alpha}$ in amounts of 250–1000 /ig produced a significantly ($P < 0.05$) smaller pupil in treated eyes as compared with control eyes at various times.

Aqueous Flare and Cellular Response in the Anterior Chamber

Cellular response in the anterior chamber was not observed under slit-lamp examination in any of these animals at any time after the topical application of PGF$_{2\alpha}$.

Some aqueous flare was observed in the anterior chamber of the treated eye of most rabbits at 0.5–5 hr after the topical application of 50 /ig or 100 /ig of PGF$_{2\alpha}$ (Fig. 7). Topical application of PGF$_{2\alpha}$ in amounts of 50 /ig at 1–3 hr ($P < 0.05$) and 100 /ig at 0.5–6 hr ($P < 0.005$) produced a significant aqueous flare in treated eyes as compared with control eyes.

Aqueous flare was observed in the anterior chamber of the treated eye of some cats at 2–6 hr after 500 /ig, 750 /ig, or 1000 /ig of PGF$_{2\alpha}$ administration (Fig. 8). This was significant ($P < 0.05$) as compared with control eyes at 3 hr after 500–1000 /ig of PGF$_{2\alpha}$ application.

Aqueous flare was not observed in any of the eyes of monkeys at any time after the topical application of 250–1000 /ig of PGF$_{2\alpha}$.

Outflow Facility

In 21 cats, 2 hr after a topical dose of 750 /ig of PGF$_{2\alpha}$, the intraocular pressure was ($P < 0.001$) reduced significantly in treated eyes as compared with baseline values and control eyes, and the mean outflow facility was increased significantly ($P < 0.001$) 48 ± 12% as compared with control eyes. In the control eyes, the outflow facility was not significantly altered, as compared with baseline values.

In monkey eyes 4 hr after administration of 500 /ig of PGF$_{2\alpha}$, the intraocular pressure was significantly reduced as compared with baseline values ($P < 0.005$).
and to control eyes ($P < 0.001$), and the outflow facility was increased significantly as compared with baseline values ($P < 0.05$) and control eyes ($P < 0.025$) (Table 1). In the control eyes, \( \text{PGF}_2 \alpha \) did not produce a contralateral alteration of outflow facility.

**Aqueous Flow**

Values of aqueous humor flow were not ($P > 0.4$) changed significantly by unilateral administration of 750 \( \mu \text{g} \) \( \text{PGF}_2 \alpha \) in cats and 500 \( \mu \text{g} \) \( \text{PGF}_2 \alpha \) in monkeys. Baseline aqueous humor flow (mean \( \mu l/min \pm SE \)) in treated and control eyes, respectively, was 22.5 ± 2.1 and 22.7 ± 3.6 in nine cats. Two hours after unilateral administration of 750 \( \mu \text{g} \) \( \text{PGF}_2 \alpha \) aqueous humor flow in 14 cats was similar in the treated eyes, 18.7 ± 1.6, and control eyes, 20.9 ± 1.7. Aqueous humor flow in the treated eyes of 10 monkeys was 1.9 ± 0.1 prior to and 1.8 ± 0.1, 4 hr after treatment and was 1.9 ± 0.1 prior to and 1.8 ± 0.1 after diluent in control eyes.

**Aqueous Humor Protein**

The protein level in the aqueous humor of the treated eyes of 11 cats, 2.02 ± 0.33 mg/ml, 2 hr after 750 \( \mu \text{g} \) of \( \text{PGF}_2 \alpha \), was significantly ($P < 0.001$) higher than that of the control eyes, 0.45 ± 0.07 mg/ml.

**Discussion**

The results presented here show that topical application of \( \text{PGF}_2 \alpha \) can reduce effectively intraocular pressure in rabbits, cats, and cynomolgus monkeys. There are, however, significant species differences. Rabbit and cynomolgus monkey eyes have a similar tendency to an initial hypertension before the onset of \( \text{PGF}_2 \alpha \)-induced hypotension. Cynomolgus monkey eyes are less sensitive than rabbit eyes to the hypertensive effects of topically administered \( \text{PGF}_2 \alpha \). No initial hypertension occurs in the eyes of cats after the topical application of \( \text{PGF}_2 \alpha \) in doses that are highly effective in reducing intraocular pressure. Moreover, the duration of intraocular pressure reduction that follows topical \( \text{PGF}_2 \alpha \) application is much longer in cat or monkey eyes than that in the eyes of rabbits. Cat eyes are clearly more sensitive to the hypertensive effects of \( \text{PGF}_2 \alpha \) than the eyes of rabbits and monkeys. Our results are similar to previously reported findings on the effects of topically applied PGs on the eyes of rabbits, cats, and monkeys. These species differences in the duration of the hypertensive effect of PGs may arise from differences between the ocular pharmacokinetics of PGs in these species.

\( \text{PGF}_2 \alpha \) reduces intraocular pressure in various species of monkeys. Previous experiments show that topical application of a single dose of 1000 \( \mu \text{g} \) \( \text{PGF}_2 \alpha \) onto the cornea of five, trained, owl monkeys produces a prolonged and highly significant ocular hypotony. The intraocular pressure of the treated eye was 4.7 ± 0.9 mmHg below that of the control eye 18 to 24 hr after \( \text{PGF}_2 \alpha \) application and remained significantly reduced for over 72 hr. Topical application of either \( \text{PGF}_2 \alpha \) or \( \text{PGE}_2 \) to the eyes of rhesus monkeys also causes significant dose-dependent reduction in intraocular pressure. The present experiments indicate that topical application of 500 \( \mu \text{g} \) of \( \text{PGF}_2 \alpha \), administered to the eyes of cynomolgus monkeys causes significant reduction in intraocular pressure at 3–24 hr after application. The maximum decrease of 4 mmHg below

**Table 1.** The effect of 500 \( \mu \text{g} \) \( \text{PGF}_2 \alpha \) on the outflow facility of 20 monkeys

<table>
<thead>
<tr>
<th>Treatment</th>
<th>( \text{Intraocular pressure} ) Mean ± SE (mmHg)</th>
<th>( \text{Outflow facility} ) Mean ± SE (( \mu \text{l}/\text{min/mmHg} ))</th>
</tr>
</thead>
<tbody>
<tr>
<td>PGF2( \alpha )</td>
<td>18.3 ± 0.4 14.9 ± 0.8*</td>
<td>0.48 ± 0.03 0.60 ± 0.04*</td>
</tr>
<tr>
<td>Control</td>
<td>17.8 ± 0.3 17.7 ± 0.8</td>
<td>0.50 ± 0.03 0.49 ± 0.04</td>
</tr>
</tbody>
</table>

* Significantly different as compared with \( \text{PGF}_2 \alpha \) treatment at 4 hr, paired t-test.

† Significantly different as compared with control eyes at 4 hr, paired t-test.
the intraocular pressure of the control eye is produced at 4 hr after 500 μg of PGF2α. However, the brief ocular hypertensive phase induced by PGF2α in various monkey species must be noted also.

In our studies, the sensitivity of cat eyes to PGF2α induced miosis is noteworthy. This finding is consistent with previous observations. No miosis is reported in rhesus monkey eyes after the topical application of 100 μg, 500 μg, or 1000 μg of PGF2α. However, our experiments show that in cynomolagus monkey, there is a small decrease in pupillary diameter in the treated eyes. This may reflect species difference. The increase in pupillary size in the control eyes remains unexplained.

Although rabbit and monkey eyes are somewhat similar with respect to the PG-induced intraocular pressure change, these two species are strikingly different with respect to their sensitivity to the generally assumed, primary, pathophysiologic effect of PGs, i.e., the breakdown of the blood–aqueous barrier. Our findings are consistent with previous observations in rabbit and cynomolgus monkey eyes. Flare was not observed previously under careful slit-lamp examination in any cats after the topical application of up to 1000 μg of PGF2α. However, in our experiments, some flare was observed in the anterior chamber of most cats 3 hr after the topical application of 500 μg, 750 μg, or 1000 μg of PGF2α. Daily or twice daily, PGE2 application to cat eyes was shown to maintain a reduced intraocular pressure for several months without causing substantial flare or cellular response and a comparison of the treated and control eyes revealed no other side effects.

To our knowledge, our study is the first tonographic and fluorophotometric analysis of the effect of PGF2α. Previous tonographic studies of the action of prostaglandins have been during the time of intraocular pressure elevation. Kass and Podos note a statistically significant increase in outflow facility after topical PGE1 in rabbits, suggesting that increased aqueous production rather than increased resistance to outflow is the cause of the intraocular pressure elevation. Camras, Bito, and Eakins report that the gross outflow resistance, measured by constant-rate infusion of the PG-treated eye is 40 to 50% of that of the contralateral control eye, in an experiment done in the hypotonic phase of eyes of rabbits after topical application of 50 μg of PGE2. They suggest that the reduction in intraocular pressure could not be due to an alteration in secretory mechanisms or pseudofacility, and, hence, must be attributed primarily to a reduction in true outflow resistance. One point we sought to clarify was the mechanism of the intraocular pressure reduction due to PGF2α. Our tonographic results demonstrate a significant mean increase in outflow facility when intraocular pressure is reduced after PGF2α therapy in cats and monkeys. However, the changes in outflow facility do not account for the total observed reduction in intraocular pressure. In our studies of the ocular hypotensive phase, one may calculate from the Goldmann flow equation, F = C (Po-Pe), that in monkeys there is a 24% reduction of aqueous flow at 4 hr after PGF2α, assuming PGF2α has no effect on episcleral venous pressure. However, direct measurements of aqueous humor formation by fluorophotometry do not show a significant (P > 0.8) change in flow in monkeys after PGF2α administration. No apparent change in aqueous flow could be accounted for by an actual decrease in flow but with a small increase in permeability in the presence of breakdown of the blood–aqueous barrier. A possible explanation of the decreased intraocular pressure due to PGF2α, in addition to an increase of pressure sensitive outflow, is an increase in uveoscleral outflow. Other mechanisms may be possible.

Absolute, tonographic results in cat eyes as calculated from the 1955 Friedenwald human tables are probably not valid. Our normal values of outflow facility are much lower than those determined by Eakins. Comparison of treated and control eyes appeared precise. Thus, the increase in outflow facility that we are reporting in cats is in percent change. Our baseline tonographic results in normal monkey eyes are similar to those reported by Kaufman and Barany, who used a two-level constant pressure perfusion technique. Three hours after the topical treatment with pilocarpine in our unpublished tonographic studies in monkeys, we find an increase of outflow facility, from 0.53 ± 0.09 μl/min/mmHg to 0.72 ± 0.08 μl/min/mmHg (Mean ± SD), similar to what they reported. Our baseline rates of aqueous flow are similar to previously reported values for cats and monkeys. However, fluorophotometric techniques to measure aqueous flow may not be valid in eyes with breakdown of the blood–aqueous barrier. PGF2α produces aqueous flare in cats but not monkeys in our study. Our findings of a small significant increase in aqueous humor protein concentration after topical PGF2α in intact eyes of cats provides evidence for some breakdown of the blood–aqueous barrier. The effect of PGF2α on aqueous humor protein in monkeys is being studied.

We also find a significant reduction in intraocular pressure of the fellow eye of cats over the baseline level when treated in one eye with PGF2α. PGE, administered to one eye, is reported to elevate the intraocular pressure of the fellow eye of rabbits. The consensual response may be due to the transfer of prostaglandin from the treated to the untreated eye via the bloodstream. However, evidence of the prompt breakdown of E prostaglandins by circulating enzymes and in the lungs makes this explanation unlikely.
ations may account for these results as well as the variations in baseline pressures in the various species. Other consensual effects cannot be ruled out.

**Key words:** prostaglandin F$_{2\alpha}$, intraocular pressure, outflow facility, aqueous humor flow, uveoscleral outflow.

**References**

7. Stern FA and Bito LZ: Comparison of the hypotensive and other ocular effects of prostaglandins E$_1$ and F$_{2\alpha}$ on cat and rhesus monkey eyes. Invest Ophthalmol Vis Sci 22:388, 1982.