The Anti-inflammatory Effects of Curcuma longa and Berberis aristata in Endotoxin-Induced Uveitis in Rabbits

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PURPOSE. To investigate the anti-inflammatory effect of topical application of Curcuma longa (C. longa) and Berberis aristata (B. aristata) aqueous extracts on experimental uveitis in the rabbit.

METHODS. Anterior uveitis was induced in rabbits by intravitreal injection of lipopolysaccharide from Escherichia coli after pretreatment with C. longa and B. aristata aqueous extracts. Subsequently, the anti-inflammatory activity of C. longa and B. aristata was evaluated by grading the clinical signs and histopathologic changes and estimating the inflammatory cell count, protein, and TNF-α levels in the aqueous humor.

RESULTS. The anterior segment inflammation in the control group was significantly higher than in both the extract-treated groups, as observed by clinical and histopathologic grading. The inflammatory cell count in the control group was 30.75 ± 7.34 × 105 cells/mL, whereas it was 2.39 ± 0.59 × 105 (P < 0.001 vs. control) and 11.56 ± 2.44 × 105 (P = 0.001 vs. control) cells/mL in the C. longa- and B. aristata-treated groups, respectively. The protein content of the aqueous humor was 18.14 ± 4.98, 3.16 ± 0.55 (P < 0.001 vs. control), and 8.24 ± 1.42 (P < 0.01 vs. control) mg/mL in the control, C. longa-, and B. aristata-treated groups, respectively. The aqueous TNF-α level in the control group was 976.29 ± 66.38 pg/mL and was 311.96 ± 28.50 (P < 0.0001 vs. control) and 654.09 ± 47.66 (P < 0.001 vs. control) pg/mL in the C. longa- and B. aristata-treated groups, respectively.


The animal model of endotoxin-induced uveitis has often been used to represent acute anterior uveitis in humans. Lipopolysaccharide (LPS), a glycolipid from the outer cell membranes of Gram-negative bacteria is a proinflammatory component and is used to induce uveitis in animals by a systemic or ocular route.1,2 Exposure to LPS induces expression of various mediators of inflammation such as tumor necrosis factor (TNF)-α and other cytokines.3–5 Increased expression of inflammatory mediators contributes to the development of anterior uveitis, with the breakdown of the blood-ocular barrier leading to infiltration of the aqueous humor with inflammatory cells and the leakage of protein.6 Experimental uveitis peaks at 24 hours after LPS injection and gradually subsides over the next 24 hours.

Curcuma longa (C. longa) and Berberis aristata (B. aristata) are two among the many herbs that have been widely used in traditional medicine for centuries. Turmeric, the powdered form of the rhizome of C. longa is rich in curcuminoids. Among the curcuminoids, curcumin is the major phenolic component in the rhizomes of C. longa. The roots of B. aristata contain significant amounts of the isoquinoline alkaloid berberine. Both the herbs are well known for their anti-inflammatory activity. The aqueous extract of the roots of B. aristata (500–1000 mg/kg) was found to have a significant anti-inflammatory effect in rats with carrageenan-induced paw edema; the effect was comparable to that of 10 mg diclofenac sodium.7 The alkaloid berberine from B. aristata has antibacterial effects and has been found to be useful in cases of trachoma.8 Berberine has also been found to be effective in experimental herpetic uveitis.9 Berberine was also shown to abolish acetaldehyde-induced NF-κB activity and cytokine production in HepG2 cells in a dose-dependent manner.10 Extracts of C. longa have also exhibited anti-inflammatory effects in standard animal models used for testing anti-inflammatory activity.11–13 Curcumin, a major chemical constituent of C. longa was found to inhibit leukotriene formation in rat peritoneal polymorphonuclear neutrophils.14 Possible benefits of oral curcumin supplementation have been observed in cases of chronic anterior uveitis.15 As in the case of berberine, curcumin exhibits an ability to suppress NF-κB activity and cyto- kine production in experimental acute liver injury.16 In view of the potent anti-inflammatory activity of C. longa and B. aristata, the present study was designed to investigate the effect of topical application of their aqueous extracts in endotoxin-induced uveitis.

MATERIALS AND METHODS

All procedures were conducted according to the ARVO Statement for the Use of Animals in Ophthalmic and Vision Research. New Zealand White rabbits weighing 2.5 to 3 kg were obtained from the animal house facility at Delhi Institute of Pharmaceutical Sciences and Research. The aqueous extracts of C. longa (rhizome) and B. aristata (roots) were provided by Promed Exports Pvt Ltd. (New Delhi, India) and were authenticated by high-performance thin-layer chromatography (HPTLC) fingerprinting. The powdered extracts were dissolved in 0.25% hydroxy propyl methylcellulose and filtered with 0.22-μm filters (Millipore, Billerica, MA). The filtrate was stored at −4°C in sterile sealed vials until further use.

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Study Design
The rabbits were divided into three groups of four rabbits each (eight eyes). The rabbits in groups 1 and 2 were topically instilled with 0.1% \textit{C. longa} and 2% \textit{B. aristata} aqueous extract, respectively, while group 3 (control group) received the vehicle. All instillations were performed three times a day for 3 days before induction of experimental uveitis and continued for 3 days after induction. Twenty-four hours after the induction of experimental uveitis, aqueous humor sampling was performed for cell count, protein, and TNF-\alpha estimation. The rabbits were reexamined 72 hours after the endotoxin injection for clinical signs of uveitis and then were killed and the eyes enucleated for histopathologic examination.

Rabbit Model of Experimental Uveitis
The endotoxin used to induce uveitis was lipopolysaccharide (LPS) from \textit{Escherichia coli} (Sigma-Aldrich, St. Louis, MO). The endotoxin was injected intravitreally in both eyes. One drop of proparacaine HCl (0.5%) solution was used for topical anesthesia. After the upper lid was retracted, 20 \mu L (100 ng) of the endotoxin solution was injected intravitreally at the 12 o’clock position, 3 to 4 mm posterior to the limbus with a 30-gauge needle. Preliminary dose-response studies were undertaken, and 100 ng was found to be the optimal dose for induction of uveitis in rabbits. Subsequently, the eyes were examined for clinical grading, and aqueous humor was tapped for inflammatory cell count, protein, and TNF-\alpha estimation. The person performing aqueous humor sampling, cell count, protein, and TNF-\alpha estimation was masked to the treatment given. The rabbits were killed after clinical assessment for signs of uveitis, 72 hours after experimental induction of uveitis.

Clinical Grading of Ocular Inflammation
The clinical signs of ocular inflammation were graded on a scale of 0 to 4 according to the scoring system described by Ruiz-Moreno et al. The grading scale was as follows: no inflammatory reaction, 0; discrete inflammatory reaction, 1; moderate dilation of the iris and conjunctival vessels, 2; intense iridal hyperemia, with flare in the anterior chamber, 3; and the same clinical signs as grade 3 plus the presence of fibrinoid exudation in the pupillary area, with intense flare in the anterior chamber, 4.

The grading of clinical signs of uveitis was performed at 24 and 72 hours after intravitreal injection of endotoxin.

Aqueous Humor Sampling
The animals were anesthetized with ketamine (20 – 40 mg/kg) and xylazine (1–2 mg/kg) by intramuscular injection in the hind limb. Proparacaine HCl (0.5%) solution was used topically to supplement the general anesthesia. During preliminary studies, attempts at aqueous humor (AH) sampling under topical anesthesia were found to be very painful and therefore further AH sampling was performed under general anesthesia. AH (150–200 \mu L) was aspirated from the anterior chamber with a 30-gauge disposable insulin syringe, and care was taken not to injure the iris or lens during the procedure. Each aqueous humor sample was diluted 50-fold with PBS.

Total Cell Count
The AH sample was suspended in an equal volume of Turk’s stain solution and cell counting was performed on a hemocytometer under light microscope. The cell count per field (each field considered equal to 0.1 mL) was estimated as an average of counts in four fields per sample. Subsequently, the number of cells per milliliter of AH was calculated.

Total Protein Estimation
Protein estimation in AH samples was performed according to the method described by Lowry et al. Briefly, 10 \mu L of AH sample was diluted with 990 \mu L of 1 N NaOH and reacted with 5 mL of copper reagent. After 10 minutes, 0.5 mL of Folin’s reagent was added and vortexed, and the samples were kept in the dark for 30 minutes. Absorbance was recorded with a spectrophotometer at 620 nm. Bovine serum albumin (BSA) was taken as a protein standard to calculate the protein content of the sample. All estimations were performed in duplicate.

The cell count and protein estimation were performed on the day of AH sampling.

**FIGURE 1.** Clinical signs of anterior uveitis in control and treated groups. (A) Just before intravitreal endotoxin injection. (B) Twenty-four and (C) 72 hours after intravitreal endotoxin injection.
TNF-α Estimation

TNF-α levels in AH were estimated by using a commercially available enzyme-linked immunosorbent assay (ELISA) kit from Diacclone (Besançon, France), per the manufacturer’s instructions. All estimations were performed in duplicate.

Histopathologic Examination

The rabbits’ eyes were enucleated 72 hours after intravitreal injection of endotoxin and fixed in 10% formaldehyde. The sections (6–8 μm thick) were stained with hematoxylin and eosin, and the structures in the immediate vicinity of endotoxin injection (i.e., anterior chamber, posterior chamber, and ciliary body) were evaluated for severity of vasodilatation, edema, and inflammatory cell infiltration. The inflammatory changes were graded on a scale of 0 to 5: none, 0; minimum, 1; discrete, 2; moderate, 3; severe, 4; and intense, 5.

RESULTS

Clinical Signs of Uveitis

The rabbits in the control group showed significantly higher grades of ocular inflammation than did the C. longa- and B. aristata–treated groups (Fig. 1). At 24 hours after intravitreal injection of endotoxin, the vehicle-treated control group had a mean score of 2.75 ± 0.56 (range, 2–4), whereas the C. longa- and B. aristata–treated groups showed a mean score of 1.13 ± 0.41 (range, 0–2) and 2.00 ± 0.41 (range, 1–3), respectively, at the same time point (Fig. 2). Clinical signs of uveitis observed at 72 hours after endotoxin injection are shown in Table 1.

Total Cell Count

C. longa- and B. aristata–treated groups showed a 92.42% and 62.40% inhibition of inflammatory cell infiltration, respectively, compared with that in the control group. In the control group, the total inflammatory cell count in the aqueous humor 24 hours after intravitreal injection of endotoxin was 30.75 ± 7.35 × 10³ cells/mL (mean ± SD, n = 8). The rabbits pre-treated with C. longa and B. aristata showed significantly lower inflammatory cell counts (2.39 ± 0.59 × 10³ cells/mL, P < 0.001 vs. control; and 11.56 ± 2.44 × 10² cells/mL, P = 0.001 vs. control) in the aqueous humor obtained at the same time point. In addition, a significantly lower inflammatory cell infiltration was observed in the C. longa–treated group than in the B. aristata–treated group (P < 0.001; Fig. 3).

Histopathologic Evaluation

The histopathologic examination of all eight rabbit eyes in the extract-treated groups showed significantly lower values in the extract-treated groups. In the C. longa–treated group the protein concentration was 3.16 ± 0.55 mg/mL compared with 8.24 ± 1.42 and 18.14 ± 4.98 mg/mL in the B. aristata–treated and control groups, respectively. The protein concentration in the C. longa–treated group was significantly lower than in the B. aristata–treated group (P < 0.001) and control group (P < 0.001). The aqueous humor protein concentration in the B. aristata–treated group was also significantly lower than in the control group (P < 0.01; Fig. 4).

Total Protein

Total proteins as estimated 24 hours after intravitreal injection of endotoxin showed significantly lower values in the extract-treated groups. In the C. longa–treated group the protein concentration was 3.16 ± 0.55 mg/mL compared with 8.24 ± 1.42 and 18.14 ± 4.98 mg/mL in the B. aristata–treated and control groups, respectively. The protein concentration in the C. longa–treated group was significantly lower than in the B. aristata–treated group (P < 0.001; Fig. 5).

Histopathologic and Clinical Score of Rabbit Eyes 72 Hours after Intravitreal Injection of Endotoxin

<table>
<thead>
<tr>
<th>Groups</th>
<th>Histopathologic Score</th>
<th>Clinical Score</th>
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<tbody>
<tr>
<td>Control</td>
<td>4.38 ± 0.41</td>
<td>2.75 ± 0.56</td>
</tr>
<tr>
<td>C. longa</td>
<td>1.13 ± 0.54†</td>
<td>0.75 ± 0.25†</td>
</tr>
<tr>
<td>B. aristata</td>
<td>2.13 ± 0.41*</td>
<td>1.63 ± 0.41†</td>
</tr>
</tbody>
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All values are group mean ± SD.
* P < 0.001 vs. control.
† P < 0.05 vs. B. aristata group.
‡ P < 0.01 vs. control.

FIGURE 2. The effect of topical instillation of aqueous extract of C. longa (CL; 0.1%) and B. aristata (BA; 2%) on the development of inflammatory signs in rabbits with experimental uveitis. The grading of clinical signs of uveitis was done 24 hours after intravitreal injection. All data are expressed as the mean ± SD. *P < 0.01 vs. control; **P < 0.05 vs. control (n = 8).

FIGURE 3. The effect of topical instillation of aqueous extract of C. longa (CL; 0.1%) and B. aristata (BA; 2%) on inflammatory cell infiltration in rabbits with experimental uveitis. The aqueous humor was collected 24 hours after intravitreal injection of endotoxin. All values are expressed as the mean ± SD. *P < 0.001 vs. control and BA, **P < 0.05 versus control (n = 8).
longa–treated group, only one eye was completely free of inflammation, whereas others showed mild edema, vasodilatation, and inflammatory cell infiltration, mainly in the posterior chamber. The mean histopathologic score in the C. longa–treated group was 1.13 ± 0.54 (range, 0–2). In B. aristata, seven eyes showed mild-to-moderate inflammatory changes, one eye showed very minimal inflammatory changes, and none was found to be completely free of inflammatory changes. The mean histopathologic grade in B. aristata–treated group was 2.13 ± 0.41 (range, 1–3). The statistical analysis of the histopathologic score showed a significantly higher degree of inflammation in the control group than in the C. longa– and B. aristata–treated groups (P < 0.001). A significantly higher degree of inflammation was observed in the B. aristata–treated group than in the C. longa–treated group (P < 0.05; Table 1, Fig. 6).

DISCUSSION

The results of the present study clearly demonstrated the protective effects of the topical application of two herbs in endotoxin-induced uveitis. The animals in the treated groups showed not only significantly reduced severity of clinical signs and histopathologic changes of uveitis but also significant reduction in aqueous humor levels of inflammatory cells, protein contents, and TNF-α compared with the control group.

The endotoxin-induced model used in this study is not an exact representation of clinical uveitis, but the inflammatory response and cytokine production in response to endotoxin closely resembles the acute phase of clinical uveitis. The endotoxin-induced inflammation can be observed during antibiotic therapy for ocular infections. The antibiotics, while destroying the bacteria, also release the LPS from their cell walls, thereby leading to an inflammatory response. The drugs found effective in endotoxin-induced uveitis may be effective in the treatment of ocular and systemic Gram-negative bacterial infections.

After intravitreal injection, the strong biological activity of the endotoxin initiates the inflammatory vascular and cellular responses. Increased production and release of monocytes, macrophages, and other inflammatory cells is associated with the release of potent inflammatory mediators in the aqueous
humor. One of these is TNF-α, which appears to be essential in the development of endotoxin-induced inflammation.2,3 Besides the chemical mediators of inflammation, cell adhesion molecules play a vital role in leukocyte adherence to vascular endothelial cells during the early acute phase of inflammation and contribute to the leakage of protein.

The drugs used in our study have demonstrated a significant anti-inflammatory effect in other studies,7–9,11–15 including improvement of chronic uveitis by oral supplementation.15 The therapeutic response to topical application of aqueous extracts of C. longa and B. aristata has demonstrated for the first time the possible utility of these drugs as a topical application for the treatment of anterior uveitis. The reduced severity of inflammatory changes observed in histopathologic examination and clinical manifestations in the inflamed eye was the result of significant inhibition of vascular and cellular inflammatory responses. The release of chemical mediators of inflammation is also suppressed secondary to inhibition of the cellular response. The suppression of vascular and cellular inflammatory responses by herbal extracts was evidenced by significantly low levels of inflammatory cells, proteins, and TNF-α levels in aqueous humor of treated animals. Significantly reduced protein levels in aqueous humor also indicates a possible inhibitory role of extract constituents on leukocyte adherence to the vascular endothelium. Direct binding to LPS of some of the extract constituents may be another mechanism of suppression of endotoxin-triggered uveitis. Inhibition of cyclooxygenase (COX)-2 may also contribute to the anti-inflammatory effect in anterior uveitis, as studies have shown selective COX 2 inhibitory activity of curcumin and berberine.21,22 Further studies of the isolated active principle components of these herbal extracts are warranted, to explore fully the mechanisms of anti-inflammatory activity in endotoxin-induced uveitis.

References