Localization and Characterization of Substance P Binding Sites in Rat and Rabbit Eyes

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Specific and high-affinity binding sites for Substance P (SP) were found in eyes from albino rabbits and rats using an in vitro autoradiographic method with 125I-Bolton Hunter SP (BHSP). Autoradiograms were generated by apposing 10–20 μm-thick cryostat eye sections to 3H-Hyperfilm or liquid emulsion and quantified by means of image-analysis procedures. Kinetic studies showed that equilibrium was reached after a 75-min incubation at room temperature. In rat retina, specific binding corresponding to approximately 90% of total binding, was reversible, of high affinity (dissociation constant [Kd], 0.13 ± 0.02 nM). Half-time for dissociation of 125I-BHSP was about 15 min. Unlabeled SP and the two neurokinins (NK) A and B competed in a concentration-dependent manner for retinal sites labeled by 125I-BHSP with the following order of potencies: SP > NKA > NKB, in agreement with a pharmacologic profile of a SP receptor site. In both species, specific binding was found in the iris sphincter muscle, choroid, and retina. In rats, detectable amounts of SP-binding sites were also expressed in the corneal epithelium and iridial stroma. Quantitative analysis of the autoradiograms revealed that the highest densities of 125I-BHSP binding sites were localized in the iris sphincter muscle in rabbits and the inner retina in rats. Invest Ophthalmol Vis Sci 32:1894–1902, 1991

Numerous neurogenic mediators released in the anterior segment of the eye by ocular injury, trauma, or noxious stimulation are known to elicit inflammatory effects, such as conjunctival hyperemia, miosis, rise in intraocular pressure, and disruption of the blood–aqueous barrier.1 If some of these biologic effects are demonstrated to be mediated directly by metabolites from the arachidonic acid cascade (released from the iris and the ciliary body),2 it is now widely recognized that neural pathways also participate in the initiation of such inflammatory events by releasing neuropeptides from sensory afferent nerves in the uveal tract.3 Substance P (SP), an undecapeptide isolated from intestine in 1931,4 was first proposed as a neurogenic mediator of antidromic vasodilation and plasma extravasation at the peripheral level5 and as a major component in the neurogenic ocular injury responses.6,7 Later, other biologically active substances, such as calcitonin gene-related peptide (CGRP) or cholecystokinin, have also been identified in ocular sensory structures8,9 and shown to play a functional role in the neurogenic inflammation (particularly blood–aqueous barrier breakdown for CGRP).10 However SP involvement in neurogenic inflammation is suggested by several pieces of evidence. First, nerve endings with immunoreactivity to SP are found in the uvea of several species, including humans,11 mainly in close association with the sphincter muscle of the iris and the smooth blood vasculature in the ciliary body. Second intracameral administration of SP induces a dose-dependent, nonmuscarinic pupil constriction associated with aqueous flare and an increase in intraocular pressure in the rabbit eye.6 Third electrical stimulation of the trigeminal ganglion or intracameral administration of capsaicin (both responsible for SP release in the anterior segment) are able to mimic SP-induced miosis.6,12,13 Fourth, (D-Pro2, D-Trp7-9)-SP, a SP antagonist, counteracts this phenomenon in rabbits and could therefore inhibit the ocular inflammatory response to laser iridial burns.14 Biochemical and immunohistochemical studies have localized SP in various vertebrate retinas.15 Cellular expression of SP-encoding mRNA was found recently in the rat retina using RNA blot and in situ hybridization.16 Although physiologic studies indi-
cate that SP has a neuromodulator action on ganglion cells in fish and in dopamine release from the retina in the rat, the role of SP immunoreactive neurons in the processing of visual information is not yet fully understood.

The presence of ocular SP binding sites was suggested previously by conventional binding techniques using membranes obtained from rat and bovine retina or bovine and rabbit iris, but the precise distribution of these receptor sites has not yet been investigated extensively. One autoradiographic report briefly mentioned the presence of binding sites in the rat retina, but no quantitative data or information on the pharmacologic profile of SP binding were presented. It is important to localize these binding sites since it is believed that most SP biologic actions are receptor-mediated. We therefore characterized and determined the anatomic localization of SP binding sites in rat and rabbit eyes using quantitative in vitro autoradiographic methods. We have used Bolton-Hunter SP (125I-BHSP; Amersham, les Ulis, France), a radiolabeled analogue of the tachykinin which has been extensively used in other organs to label SP receptors.

Materials and Methods

Tissue Preparation

New Zealand albino rabbits (weighing 3–3.5 kg) were killed by injection of a lethal dose of sodium pentobarbital and Wistar rats (weighing 200–250 g), by decapitation. The eyes were removed, immersed in Tissue Tek medium (Miles Scientific, Naperville, IL), frozen in isopentane cooled in liquid nitrogen and stored at -80°C. Just before sectioning, the tissues were warmed to -20°C, and sections (20-μm thick) were cut with a cryostat (Bright), thaw-mounted onto gelatin-coated glass slides, and stored at -80°C until use. Before incubations, sections were allowed to thaw at room temperature. All investigations described in this paper were done in accordance with the ARVO Resolution on the Use of Animals in Research.

Binding Conditions

Preliminary experiments showed that preincubation was found to increase specific binding. Thus, slides were washed in a preincubation medium (Tris-HCl 50 mM, pH 7.4, containing 0.2 g/l of bovine serum albumin) at room temperature for 15 min before incubation with radioligand. After the washing step, the slides were then incubated at room temperature in a solution of 65 pM 125I-BHSP (2000 Ci/mmol) (Bolton-Hunter is 3-(p-hydroxy-m-(125I)iodo-phenyl)propionyl) in 50 mM Tris-HCl, pH 7.4, containing 10 mM MgCl2, 2 g/l of bovine serum albumin, 40 mg/l of bacitracin, 5 mg/l of leupeptin, and 4 mg/l of bestatin. Nonspecific binding was determined on alternate sections in the presence of 10 μM unlabeled peptide (SP) added in the incubation medium. The specificity of the binding was studied by incubating sections with increasing concentrations of peptides related to SP, such as senktide, peptide, spantide, SP methylester, neurokinin A (NKA), neurokinin B (NKB), and synthetic fragments of SP (SP1-4 and SP4-11). Bacitracin, leupeptin, and bestatin were obtained from Sigma (St. Louis, MO) and SP-related peptides from Bachem (Bubendorf, Switzerland). After incubation with the radioligand, the sections were rinsed four times for 1 min each in the precubation buffer at 4°C, dipped for 20 sec into distilled water, and quickly dried using a stream of cold air for autoradiography.

Autoradiographic Experiments

The sections were preincubated as described and incubated with 65 pM 125I-BHSP for 90 min at room temperature. After washing, the dry slides were stored in a Kodak X-ray cassette (Rochester, NY) in tight apposition to a tritium-sensitive film (3H-Hyperfilm; Amersham) and allowed to expose for 1 week in darkness. After exposure, the films were developed in Kodak D19 for 3 min and fixed. To identify the localization of the binding sites, the eye sections were counterstained with hematoxylin and eosin and covered slipped with Fluca (Chemika, Buchs, Switzerland).

To investigate precisely the cellular localization of SP binding sites, an autoradiographic technique was used at light microscopic resolution. Briefly, after incubation and washings, selected slides were treated with a 30-min bath of 4% glutaraldehyde at 4°C to fix covalently the radioligand to its binding site, defatted in several baths of increasing concentrations of alcohol (75–100%) and xylene, and then dipped into liquid nuclear emulsion (LM1; Amersham). Preliminary experiments showed that this treatment did not significantly alter 125I-BHSP binding. After an exposure period of 10 days in darkness, the emulsion dipped microautoradiograms were developed and fixed as described. Corresponding sections were counterstained and examined under a light microscope.

Data Analysis

Film microautoradiograms were analyzed by computer-based densitometry. The optical density of the autoradiogram was quantified by means of an image.
analyzer (BIOCOM RAG 200, les Ulis, France). Briefly, autoradiograms were digitized, and each grain density was assigned a relative optical density value. For maximal binding capacity determination, these relative optical density values were then converted to corresponding commercial $^{125}$I Amersham standards, and the results were expressed in fmol/mg tissue equivalent. Amersham’s microscales are supplied as slices of several layers of polymer containing a range of increasing $^{125}$I concentrations. Since tissue sections are coexposed with iodinated plastic standards, we can do quantitative densitometry using a computer-assisted system. Tissue-equivalent values were provided based on calibration using intact brain gray matter. In all cases, specific binding was defined by subtracting nonspecific binding (obtained in the presence of $10^{-6}$ M of unlabeled SP) from total binding. Classic computer analysis was used for the biochemical determination of the binding parameters (association constant [Kd] and concentrations which inhibit 50% of the specific binding [IC$_{50}$]) on rat retina sections. The kinetics of $^{125}$I-BHSP binding was assumed to follow a bimolecular association model (second-order kinetics).

Results
Characterization of $^{125}$I-BHSP Binding

Kinetic analysis done by densitometry on rat retinal sections indicated that the association of $^{125}$I-BHSP resulted in a time-dependent increase in binding (Fig. 1). At room temperature, specific binding reached a plateau in approximately 75 min; nonspecific binding was not significantly increased. About 90% of total binding was specific at 75 min. A routine incubation time of 90 min was then adopted for all subsequent experiments. At equilibrium, bound $^{125}$I-BHSP could be dissociated specifically from its binding sites by incubating sections with an excess ($10^{-5}$ M) of unlabeled peptide (Fig. 1). The half-time for dissociation of $^{125}$I-BHSP was about 15 min.

We investigated competition studies between iodinated SP and increasing concentrations of unlabeled peptide (Fig. 2). These results indicated that SP inhibited $^{125}$I-BHSP competitively and with very high affinity. In the range of concentrations studied ($10^{-11}$ to $10^{-6}$ M), IC$_{50}$ values were 0.17 nM. Curve analysis obtained (obtained by regression lines computerized by means of the least-squares method) indicated that $^{125}$I-BHSP bound to high-affinity binding sites in rat retina with an apparent Kd estimated at 0.13 ± 0.02 nM.

Competition studies with the tachykinins NKA, NKB, and analogues or fragments of SP were done in attempt to establish the type of site identified by $^{125}$I-BHSP in the rat retina. The order of potency for inhibition of binding was SP > NKA > NKB (Fig. 3, Table 1). A similar ranking was obtained when rabbit eye sections were incubated with the tachykinins at concentrations in the range of $10^{-8}$ to $10^{-5}$ M (data not shown). Displacement curves showed that specific binding...
binding of $^{125}$I-BHSP was strongly inhibited by SP$_{1-11}$ (IC$_{50}$, 0.17 ± 0.02) (Fig. 2) and to a lesser extent by SP$_{4-11}$ and NKA (IC$_{50}$ = 3.39 and 8.04 nM, respectively). On the other hand, senktide and septide were ineffective, and SP$_{1-4}$, NKB, and spantide had some potency to inhibit retinal $^{125}$I-BHSP binding (Table 1).

**Discussion**

These results demonstrate that several structures from albino rat and rabbit eyes (mainly the retina and the iris sphincter muscle) have the capacity to bind specifically a derivative analogue of SP, $^{125}$I-BHSP. The characteristics of $^{125}$I-BHSP binding in the rat retina strongly suggest that the ligand binds to SP receptors.

The apparent Kd of the high-affinity (0.13 nM) binding site identified in this study was similar to those reported in rat retina homogenates (0.2 nM), in rabbit optic sections (0.54 nM), and in human retina (0.27 nM). Recent studies have described the existence of multiple tachykinin receptors in the rat retina. The agent, SP, belongs to a group of closely
the mammalian tachykinins have been described, namely SP (or NK-1) receptors, found in both central and peripheral tissues, NKA (or NK-2) receptors in peripheral tissues, and NKB (or NK-3) receptors mainly detectable in the central nervous system. Our data suggest that 125I-BHSP binds preferentially to NK-1 receptor sites; SP is the most potent tachykinin in inhibiting 125I-BHSP binding and the order of potency of the competitors for 125I-BHSP binding was SP > NKA > NKB. The IC_50 values calculated for NKA and NKB were somewhat lower than those found for the displacement of 125I-BHSP binding in a preliminary report. Variations in the methods, including the incubation buffer and the lower ligand concentration used in our study (65 pM), may be the cause of these differences. It is unlikely that 125I-BHSP binds to NKB receptors since senktide, a highly selective NK-3 agonist, showed no activity at the micromolar level in displacing 125I-BHSP from its binding sites.
binding sites were found to be mainly associated with the iris sphincter muscle although other iridal structures were slightly labeled in the rat eye. The presence of SP was reported in sensory neurons of the anterior uvea originating from the trigeminal ganglion,33,34 and a number of investigators extensively studied the involvement of the peptide in the acute irritation response of the eye. Besides its vasodilator properties, early experiments reported that the tachykinin peptide induces an increase in intraocular pressure and moderate alterations in the blood–aqueous barrier in rabbit eyes.6,7,14,25 Subsequent studies observed that these effects were relatively weak and inconsistent and that the rise in intraocular pressure was mainly due to a miosis-induced pupillary blockade since it was abolished in part by peripheral iridectomies.12 Despite its wide distribution in the uveal tract in many species,11,23,34,36,37 SP has been proposed to be only responsible for the miotic component of the antidromic ocular injury response.12,35,38 The other parts of the inflammatory response (elevated intraocular pressure and blood–aqueous barrier breakdown) are thought to be mediated by other mechanisms; in animals pretreated with prostaglandin inhibitors, injection of SP caused only miosis.12 The release of CGRP from sensory nerves has also been shown to induce marked inflammatory effects in cat39 and rabbit40 eyes. The two neuropeptides are partially colocalized in terminal endings arising from the trigeminal cells41 and are released together into aqueous humor during the antidromic response. In addition, SP-induced miosis is potentiated by CGRP,42 suggesting biologic interactions between the two peptides. Our autoradiographic results provide essential information concerning the role of SP in the ocular anterior segment; they demonstrate the presence of high-affinity binding sites for SP associated with the iridial sphincter muscle. They may also suggest that the miotic response to SP in rabbits may be due to a direct action site. Furthermore, as described for SP binding sites in other tissues,19,25,26 affinity is encoded in the carboxy terminal of SP since SP_{1-11} and SP_{2-11} had the greatest potency in inhibiting ^{125}I-BHSP binding.

Autoradiographic visualization of ^{125}I-BHSP binding sites in the anterior segment of the eye shows a selective pattern of distribution. In both species, SP

**Table 2.** Specific ^{125}I-BHSP bound (fmol/mg tissue equivalent)

<table>
<thead>
<tr>
<th>Area</th>
<th>Specific ^{125}I-BHSP bound (fmol/mg tissue equivalent)</th>
</tr>
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<tbody>
<tr>
<td>Cornea</td>
<td>0.12 ± 0.01</td>
</tr>
<tr>
<td>Iris stroma</td>
<td>0.11 ± 0.01</td>
</tr>
<tr>
<td>Iris sphincter muscle</td>
<td>2.93 ± 0.42</td>
</tr>
<tr>
<td>Retina</td>
<td>0.95 ± 0.17</td>
</tr>
<tr>
<td>Retina</td>
<td>0.63 ± 0.07</td>
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Regional distribution of specific BHSP binding sites in the albino rat and rabbit eye. Results were obtained by transformation of optical densities as described in the text to fmol/mg tissue equivalent ± SEM and were obtained from at least 10 determinations throughout the area under examination. Nonspecific binding was subtracted from each total value, so that data are expressed as specific binding.
on the iris muscle through specific receptors. No binding sites was found in dilator or ciliary muscles, corroborating the fact that SP has no contractile effect on these tissues.\textsuperscript{38,42} One of the most interesting findings of our study was the absence of high-affinity binding sites in the rabbit iris and ciliary epithelia, known to be the major site of the blood–aqueous barrier. It is unlikely that the lack of binding sites in rabbit ciliary body may be due to occupied receptor sites since sections were washed in a preincubation medium for 15 min before incubation with \textsuperscript{125}I-BHSP. Nevertheless, we cannot exclude the possibility of the presence of lower-affinity receptors or other unknown subtypes in ciliary process. Such sites may be absent or not detectable by our autoradiographic procedures. However, these results suggest that, in the rabbit, the peptide is not directly involved in the blood–aqueous barrier disruption observed in ocular neurogenic inflammation, as suggested by its weak and inconsistent effect. In addition, we cannot exclude the possibility that SP exerts some inflammatory actions in the eye through an indirect mechanism; it has been demonstrated that SP has a wide spectrum of inflammatory properties, in particular mast cell degranulation with release of histamine\textsuperscript{44} and arachidonic acid\textsuperscript{45} (and consequently prostaglandin biosynthesis). Finally, the involvement of SP in ocular neurogenic inflammation cannot be generalized since it has been reported that the ocular response to SP was species-dependent (cat, baboon, or human iris are relatively insensitive to SP\textsuperscript{46}). This species variation is well illustrated in the our study by the fact that the overall density of iridal SP binding sites was higher in the rabbit than in the rat.

The distribution of uveal SP binding sites and SP nerve terminals was not correlated well. In our study, regions that have relatively high densities of SP binding sites, such as the iris sphincter muscle, have been reported previously to have the highest amounts of SP immunoreactivity.\textsuperscript{47} In contrast, regions such as the ciliary processes or the iridial stroma, also known to contain SP immunoreactivity, did not have detectable SP binding sites. The existence of discrepancies between the localization of receptors and the distribution of fibers or neurotransmitters has been clearly demonstrated for SP in the brain.\textsuperscript{48}

The SP binding activity in the scleral vessels (Fig. 5) may be associated with the endothelium or vascular constituents. This may be attributed to the chemotactic peptide, f-Met-Leu-Phe (MLP)\textsuperscript{49} or the phagocytosis promoting peptide, tuftsin,\textsuperscript{50} which share structural similarities with SP. Furthermore, SP has been shown to interact with the tuftsin receptors present on the macrophage and polymorphonuclear leukocyte plasma membrane.\textsuperscript{50} However, under our experimental conditions, competition studies with increasing concentrations of tuftsin (10\textsuperscript{-10} to 10\textsuperscript{-7} M) showed that \textsuperscript{125}I-BHSP binding was not inhibited by tuftsin (data not shown).

Although SP-containing neurons have been identified in the retina of mammals,\textsuperscript{51} including humans,\textsuperscript{52} very little is known about their functional role in this tissue. These neurons are present in most species in amacrine cells whose somata are located at the border of the INL and IPL (which send processes primarily to the IPL). Evidence for SP localization to ganglion cells has also been demonstrated in some species, particularly rabbits.\textsuperscript{53} The localization of SP binding sites we found was similar to the pattern of SP-immunoreactive cells previously described for this region. Neonatal monosodium glutamate treatment of rats (which induces degeneration of the inner layers of the retina) has been reported to cause a marked reduction in \textsuperscript{125}I-BHSP retinal binding.\textsuperscript{19} Our autoradiographic findings also agreed with this experiment since most of the SP binding sites identified our study were concentrated in the IPL and GCL. Several observations suggest a neuromodulator role for SP in retinal functions; it has been found to elicit \textsuperscript{3}H-dopamine release from rabbit retinas\textsuperscript{18} and to exert excitatory effects on carp cholinergic-sensitive ganglion cells activity.\textsuperscript{17} It has also been shown to stimulate the accumulation of inositol triphosphates in rabbit retinal cultures.\textsuperscript{54} The presence of SP binding sites in the inner retina is an additional indication that the peptide probably has some regulatory action in visual processing.

**Key words:** substance P, substance P receptor, autoradiography, eye

**References**


46. Unger WG and Tighe J: The response of the isolated iris


