Immunoglobulin A Antibodies Against *Pseudomonas aeruginosa* in the Tear Fluid of Contact Lens Wearers

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**Purpose.** *Pseudomonas aeruginosa* is the most important cause of contact lens-associated ulcerative keratitis, especially for those who use extended-wear lenses. Until now, the presence of specific anti-*P. aeruginosa* immunoglobulin A (IgA) antibodies in the tears of contact lens wearers has not been investigated and is the purpose of the current study.

**Methods.** The levels of specific IgA antibodies against *P. aeruginosa* and total secretory IgA (s-IgA) concentrations were measured in tears of various groups of contact lens and non-contact lens wearers using enzyme-linked immunosorbent assays. Contact lens groups were divided into the following categories: daily-wear rigid gas-permeable lenses (n = 23), daily-wear soft lenses (n = 22), extended-wear soft lenses (n = 17), and non-contact lens wearers (n = 23). As a positive control group, we tested tears obtained from patients with cystic fibrosis (n = 5) because the respiratory tract of these persons often are colonized by *P. aeruginosa*.

**Results.** The percentage of nonresponders (<15 U/ml) varied between 9% in daily-wear rigid gas-permeable contact lens users to 23% in daily-wear soft contact lens users. The percentage of nonresponders in controls was 13%. The frequency of nonresponders was not significantly different among the different groups tested. All patients with cystic fibrosis showed a very high anti-*P. aeruginosa* IgA response in their tears. When analyzing the mean anti-*P. aeruginosa* IgA response, a significantly lower level was found in extended-wear contact lens users (38 U/ml) compared to non-contact lens wearers (82 U/ml). Total s-IgA levels in the tears of the various groups tested were not significantly different.

**Conclusions.** A substantial number of persons in the population of contact lens wearers tested lack detectable IgA antibodies against *P. aeruginosa* in their tears and may be susceptible to *P. aeruginosa* keratitis if the physiological condition of their cornea is compromised. Invest Ophthalmol Vis Sci. 1996;37:2081-2088.
The subjects had worn contact lenses for at least 6 months and who have no detectable specific IgA response may be at risk for P. aeruginosa-induced keratitis when their lenses are contaminated and their corneal epithelium is compromised.

MATERIALS AND METHODS

Subjects

Tears were collected from 62 consecutive asymptomatic cosmetic contact lens wearers attending three contact lens clinics for routine ocular examination. The subjects had worn contact lenses for at least 6 months (mean wearing time, 10 years). The contact lens group was divided into the following categories:

1. Daily-wear rigid gas-permeable contact lens users (n = 23; mean age, 35 years).
2. Daily-wear soft contact lens users (n = 22; mean age, 27 years).
3. Extended-wear soft contact lens users (n = 17; mean age, 36 years).

The control group consisted of persons (n = 23; mean age, 32 years) who had never worn lenses before and who attended the clinics to have contact lenses fitted.

As a positive control group, we used tears from five patients with cystic fibrosis; three were colonized by P. aeruginosa, as shown by bacteriologic sputum cultures. Slit lamp examination documented that none of the contact lens wearers or controls had ocular disease.

The tenets of the Declaration of Helsinki were followed, and the study was approved by the human experimentation committee of the Medical Faculty of the University of Amsterdam. Informed consent was obtained from subjects before collection of tears.

Tear Collection

Tears were collected by placing a sterile cellulose sponge (Sugi, Kettenbach, Germany) in the conjunctival sac of both eyes as described earlier. Tear collection was performed between 9 AM and 11 AM at the outpatient department of the hospital. Sponges were removed after 1 minute, and tear fluid was separated from the sponges by centrifugation and stored at −20°C. In contact lens wearers, lenses were removed before tear collection.

Strains and Serotyping

P. aeruginosa was isolated from the corneal ulcer of a patient with a keratitis (reference strain 474) and serotyped (serotype P1) according to the Sanofi Diagnostics Pasteur (Marnes La Coquette, France) typing scheme, which is identical to the O1 serotype using the International Antigenic Typing Scheme. The O1 serotype is a strain commonly isolated in patients with P. aeruginosa keratitis.

Bacterial Cell Culture

P. aeruginosa was grown overnight on blood agar plates at 37°C. One colony was inoculated in 100 ml of broth medium containing 5% Bacto peptone (Difco Laboratories, Detroit, MI) and 0.25% trypticase soy broth (Difco). The culture was grown on a rotary shaker at 37°C for 18 hours, centrifuged at 7000g for 20 minutes at 4°C, washed three times with phosphate-buffered saline (PBS, 160 mM Na, 1.3 mM H2PO4, 9.2 mM HPO4, and 140 mM Cl, pH 7.4), and suspended in PBS to a concentration of 10⁶ colony forming units (CFU)/ml. The bacteria were heated at 100°C for 2 hours, divided into 1 ml aliquots, and stored at −20°C before use.

Bacterial Culture of Contact Lens Containers

Before the visits, all subjects were not informed about the contamination survey and were not instructed to bring their lens storage cases with them. After tear collection, all contact lens wearers were asked whether they had brought their lens cases. If so, lens cases were exchanged for new cases. Otherwise, they were requested to send their lens containers by mail to the research institute the next day. In total, 56 lens cases (44 of the daily-wear lens users and 12 of the extended-wear lens users) were collected. The presence of P. aeruginosa in these storage cases was investigated using a P. aeruginosa-specific broth (special peptone, casein hydrolysate with “P. aeruginosa supplement” consisting of 100 µg/ml cetrimide and 15 µg/ml sodium nalidixate). All media components were purchased from Oxoid (Basingstoke, UK). The fluid in the lens...
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containers was removed, and they were washed with culture medium to remove remaining care solution. The special P. aeruginosa bouillon was added to the containers, which were then vigorously vortexed and incubated for 24 hours at 37°C. A sample of the medium was cultured on cysteine, lactose, electrolyte-deficient plates (Oxoid) and P. aeruginosa-specific agar plates (containing cetrimide and sodium nalidixate; Oxoid) for 2 days.

**P. aeruginosa IgA Enzyme-Linked Immunosorbent Assay**

Initial experiments were performed whereby the enzyme-linked immunosorbent assay (ELISA) was compared using heat-killed (60°C for 1 hour and 100°C for 2 hours) versus freshly cultured bacteria. Killing was verified by bacterial culture after heating. Highest absorbance readings were obtained with bacteria that had been pretreated at 100°C for 2 hours. We, therefore, used heat-killed bacteria (100°C for 2 hours), which is similar to protocols used by previous investigators. A dose–response curve of the concentrations of bacteria used for coating the ELISA plates revealed a bell-shaped curve at which 1.25 × 10^7 CFU/ml was optimal.

The above-mentioned experiments led to the following protocol. Heat-killed (100°C for 2 hours) P. aeruginosa organisms (10^9 CFU/ml) were diluted 1:80 in 0.05 M carbonate-bicarbonate coating buffer (pH 9.6). ELISA plates (96 wells; #566101; Greiner, Frickenhausen, Germany) were incubated overnight with the organisms (150 µl/well) at room temperature and were washed four times with PBS the next morning. The wells subsequently were blocked with PBS containing 2% (wt/vol) bovine serum albumin (BSA) and 1 mM ethylenediaminetetraacetic acid (EDTA), pH 7.3 (blocking buffer, 150 µl/well) during 2 hours at room temperature. Plates were washed and filled with 100 µl/well of serial tear fluid dilutions (1:25 and 1:50 in blocking buffer with 0.1% (vol/vol) Tween 20) and incubated for 1 hour at 37°C. After this incubation, the wells were washed and incubated with a horseradish peroxidase-labeled rabbit anti-human IgA antibody (Dako, Copenhagen, Denmark) diluted 1:500 in PBS containing 0.1% (wt/vol) gelatin, 2% (wt/vol) normal rabbit serum, and 0.02% (vol/vol) Tween 20, pH 7.3. After a 1-hour incubation at 37°C and a washing procedure, 150 µl/well of substrate buffer was added: 0.11 M acetic acid, 0.01% (wt/vol) tetramethylbenzidine, 1% dimethylsulfoxide, and 0.03% H2O2, pH 5.4. The enzyme substrate reaction was inhibited after 10 minutes by the addition of 50 µl/well of 2 M H2SO4. The optical density was measured in a Titertek Multiscan MC (Amstelstad, Netherlands) at 450 nm. Results were expressed after subtraction of the corresponding optical density of negative control wells for each sample.

![Figure 1](https://iovs.arvojournals.org/pdfaccess.ashx?url=/data/journals/iovs/933413/ on 11/03/2018)

**Figure 1.** Standard curve of anti-Pseudomonas aeruginosa immunoglobulin A. IgA antibodies directed against P. aeruginosa were determined by enzyme-linked immunosorbent assay on a pool of tears that had been set arbitrarily to contain 1000 U/ml. Plates either were coated with P. aeruginosa (●—●) or with coating buffer (○—○) alone.

Negative controls of the ELISA included the incubation of uncoated wells with tear dilutions and the incubation of P. aeruginosa coated wells with buffer instead of tears.

Initial experiments showed that in some persons, their tears contained factors that gave a high background when incubated with the uncoated wells. Addition of normal rabbit serum to the peroxidase-labeled antihuman IgA antibody solved this problem. The ELISA readings were converted to units using a calibration curve obtained by including serial dilutions of a pool of human tears from persons with a high anti-P. aeruginosa IgA response. This pool of tears had been set arbitrarily to contain 1000 U of anti-P. aeruginosa IgA per ml. Each plate always contained its own standard calibration curve. A dose–response curve of the standard pool of tears is shown in Figure 1. A computer program (Logit; Central Laboratory, Dutch Red Cross Blood Transfusion Service, Amsterdam, Netherlands) was used to generate standard curves and to calculate anti-P. aeruginosa IgA levels in experimental samples. From the standard curve, it can be seen that reliable measurements could be obtained at tear dilutions of less than 1:1600, corresponding to 0.625 U/ml. Because experimental tear samples were tested at a dilution of 1:25, the sensitivity of our test was 15 U/ml. Persons with ELISA readings less than 15 U/ml were considered nonresponders. Analytical variation of the ELISA, including intraplate (intra-assay) and day-to-day (interassay) variations, was determined with two tear samples (10 determinations for each). The intra-assay and interassay variations of the ELISA were 2% and 7.5%, respectively.

To prove that the response in our P. aeruginosa ELISA was associated with s-IgA, the following experiment was performed. Tears were collected from a
**P. aeruginosa** responder, 25 µl was loaded onto an Fast Performance Liquid Chromatography column (SMART System; Pharmacia, Uppsala, Sweden), and tear proteins were fractionated according to their molecular weight. The first peak emerging from the column and known to contain the IgA peak was collected (total volume, 175 µl) and tested in the ELISA, together with the original tear sample; a correction was applied for the dilution (1:7) occurring during the gel filtration procedure. Both the 1:20 and 1:100 dilutions gave similar optical density readings in the ELISA. The 1:20 dilution gave a reading of 2.04 for the original tears compared to 1.97 for the isolated IgA fraction, whereas the 1:100 dilution gave an optical density reading of 1.33 (whole tears) versus 1.18 (IgA fraction).

### s-IgA ELISA

s-IgA in human tear fluid was quantitated using a sandwich ELISA; s-IgA was captured with a monoclonal antisecretory component antibody (Nordic Immunological Laboratories, Netherlands) and detected with the same peroxidase-labeled rabbit antihuman IgA antibody as described above. ELISA plates (96 wells; #566101, Greiner) were coated overnight with 0.1 mg/ml monoclonal antisecretory component antibody in PBS. After washing, the wells were incubated with serial tear dilutions (1:500, 1:1000, 1:2000) in PBS containing 0.1% (vol/vol) Tween 20. The same tear sample was used as for the detection of the anti-**P. aeruginosa** IgA response. After incubation for 1 hour at 37°C, the wells were washed, and bound s-IgA was detected by incubating (1 hour at 37°C) with a peroxidase-labeled anti-IgA diluted in 1:500 PBS containing 0.1% (vol/vol) Tween 20 and 2% (wt/vol) normal rabbit serum. The ELISA was developed using the above-mentioned substrate and quantitated as mg s-IgA/ml using a calibration curve with human colostrum s-IgA (Sigma, St. Louis, MO) as a standard.

### Western Blot Analysis

For immunoblotting, an outer membrane preparation of the bacteria was made as follows. **P. aeruginosa** was cultured as described above. Bacteria were harvested by centrifugation at 2500g for 10 minutes at 4°C. The pellet was resuspended in 7 ml of 50 mM Tris–HCl (pH 7.8). The suspension was sonicated intermittently for 5 minutes on ice. After centrifugation at 2000g for 20 minutes at 4°C, the supernatant was centrifuged in an ultracentrifuge at 45,000g for 1 hour at 4°C. The supernatant was discarded, and the pellet was resuspended in 200 µl of 2 mM Tris–HCl pH (7.8).

Sodium dodecyl sulfate–polyacrylamide gel electrophoresis was performed on the Phast System (Pharmacia) using precast 10% to 15% gradient minigels. The electrophoresis conditions and silver staining of the minigels were performed according to manufacturer's instructions. The outer membrane samples were prepared in a nonreducing sample buffer (62 mM Tris, 1 mM EDTA, 2% [wt/vol] SDS, 0.005% [wt/vol] bromophenol blue, pH 8) and had a protein content of approximately 0.2 µg/µl. After electrophoresis, separated outer membranes were transferred to Immobilon PVDF membranes (pore size 0.45 µm; Millipore, Milford, MA) by electroblotting in a semidry environment. The transfer buffer consisted of 192 mM glycine, 25 mM Tris base, 2% (vol/vol) methanol, pH 8.3, and proved to have a good transfer efficiency as assessed by silver staining of gels before and after electroblotting.

After transfer, blots were incubated for 2 hours in 2% (wt/vol) BSA in 0.5% (vol/vol) Tween 20, 10 mM EDTA, pH 7.3, in PBS buffer, to block the nonspecific binding sites. Between the blotting steps, the blots were washed in PBS for 15 minutes. Blots were cut into strips. Tears were diluted to a concentration containing 5 to 10 U anti-**P. aeruginosa** IgA/ml, as previously determined by ELISA. Each immobilon strip was incubated separately with a diluted tear sample for 1 hour at 37°C. This step was followed by incubation with 1:1000 diluted biotinylated rabbit-antihuman-IgA antibody (Dako) in 3% (wt/vol) BSA.

A streptavidin–biotin–HRP complex (StrepAB-Complex; Dako) was used to detect immunoglobulin–biotin complexes. StreptABComplex, in combination

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**TABLE 1. Immunoglobulin A Response Against *Pseudomonas aeruginosa* in Tears of Contact Lens and Non-Contact Lens Wearers (Controls) and Patients With Cystic Fibrosis**

<table>
<thead>
<tr>
<th>Responders</th>
<th>Nonresponders</th>
<th>Nonresponders (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Daily-wear RGP</td>
<td>21</td>
<td>2</td>
</tr>
<tr>
<td>Daily-wear soft</td>
<td>17</td>
<td>5</td>
</tr>
<tr>
<td>Extended-wear soft</td>
<td>14</td>
<td>3</td>
</tr>
<tr>
<td>Controls</td>
<td>20</td>
<td>3</td>
</tr>
<tr>
<td>Cystic fibrosis</td>
<td>5</td>
<td>0</td>
</tr>
</tbody>
</table>

RGP = rigid gas-permeable.

* Responders had an IgA anti-*P. aeruginosa* level above 15 U/ml.
TABLE 2. Levels of Anti-Pseudomonas aeruginosa Immunoglobulin A and Total s-IgA in Tears of Contact Lens and Non-Contact Lens Wearers (Controls)

<table>
<thead>
<tr>
<th></th>
<th>IgA Anti-P. aeruginosa (units/ml)</th>
<th>s-IgA (mg/ml)</th>
<th>Ratio</th>
</tr>
</thead>
<tbody>
<tr>
<td>Daily-wear RGP</td>
<td>67 ± 11</td>
<td>1.2 ± 0.2</td>
<td>63 ± 6</td>
</tr>
<tr>
<td>Daily-wear soft</td>
<td>52 ± 9</td>
<td>0.7 ± 0.1</td>
<td>74 ± 8</td>
</tr>
<tr>
<td>Extended-wear soft</td>
<td>38 ± 6*</td>
<td>0.6 ± 0.1</td>
<td>76 ± 10</td>
</tr>
<tr>
<td>Controls</td>
<td>82 ± 15</td>
<td>0.9 ± 0.1</td>
<td>91 ± 10</td>
</tr>
</tbody>
</table>

Values are mean ± standard error of the mean.

*P < 0.03 for the difference between non-contact lens and extended-wear soft contact lens users.

with a biotin-labeled secondary antibody, has a very high sensitivity. The streptavidin–biotin–HRP complex was visualized using diaminobenzidine–metal concentrate (Pierce, Rockford, IL). Negative controls included strips containing bacterial outer membranes; in the first step, buffer alone was used instead of tears.

**Statistical Analysis**

The level of significance of the data obtained from the various groups investigated was tested using the Mann–Whitney test.

**RESULTS**

The ELISA for the detection of IgA antibodies against *P. aeruginosa* was developed using heat-killed *P. aeruginosa* obtained from a patient with a corneal ulcer. When analyzing the anti-*P. aeruginosa* IgA response in various groups of contact lens wearers and controls, the following observations were made. The percentage of nonresponders (ELISA reading less than 15 U/ml) varied between 9% in daily-wear rigid gas-permeable contact lens users to 23% in daily-wear soft contact lens users (Table 1). The percentage of nonresponders (ELISA reading less than 15 U/ml) varied between 9% in daily-wear rigid gas-permeable contact lens users to 23% in daily-wear soft contact lens users (Table 1). The percentage of nonresponders in controls was 13%. In view of the small amount of persons tested, these data did not reach statistical significance. When analyzing the mean anti-*P. aeruginosa* IgA response in the tears of the various groups, a significantly lower response was noted in the group of extended-wear soft contact lens users compared to controls (Table 2). Controls had a mean IgA anti-*P. aeruginosa* response of 82 U/ml (range, 6.1 to 285 U/ml), whereas users of extended-wear soft contact lenses had 38 U/ml (range, 5.9 to 102 U/ml) in their tear samples. All patients with cystic fibrosis had a detectable anti-*P. aeruginosa* response, and those persons chronically infected by the bacterium (sputum culture positive for *P. aeruginosa*) had extremely high levels (Table 3).

Because the low anti-*P. aeruginosa* IgA response observed in extended-wear soft contact lens users could have resulted from a decrease in the total s-IgA level in the collected tear samples, we measured the s-IgA concentration in the same sample. Although the s-IgA level in extended-wear soft contact lens users appeared to be lower than that seen in controls, no statistically significant differences were found between the groups investigated, including the patients with cystic fibrosis (Tables 2, 3).

Because tear flow rate affects the concentration of a number of tear proteins including s-IgA, we measured the volume of tears produced per minute. The volume of collected tears ranged from 35 to 75 μl, and no significant differences in the mean tear volumes were observed among the different groups tested. Moreover, using our method of tear collection, no correlation was observed between the amount of tears collected and the s-IgA concentration.

Subsequently, the specific anti-*P. aeruginosa* IgA response was calculated for each individual sample as ELISA unit anti-*P. aeruginosa* per milligram of total

TABLE 3. Levels of Anti-Pseudomonas aeruginosa Immunoglobulin A and Total s-IgA in Tears of Patients With Cystic Fibrosis

<table>
<thead>
<tr>
<th>Patient Number</th>
<th>Age (years)</th>
<th>Sputum Culture</th>
<th>IgA Anti-P. aeruginosa (units/ml)</th>
<th>s-IgA (mg/ml)</th>
<th>Ratio</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>24</td>
<td>+</td>
<td>2060</td>
<td>0.3</td>
<td>6867</td>
</tr>
<tr>
<td>2</td>
<td>18</td>
<td>-</td>
<td>92</td>
<td>0.2</td>
<td>460</td>
</tr>
<tr>
<td>3</td>
<td>40</td>
<td>-</td>
<td>452</td>
<td>1.9</td>
<td>238</td>
</tr>
<tr>
<td>4</td>
<td>20</td>
<td>+</td>
<td>6218</td>
<td>0.6</td>
<td>10,363</td>
</tr>
<tr>
<td>5</td>
<td>24</td>
<td>+</td>
<td>28550</td>
<td>1.8</td>
<td>15,861</td>
</tr>
</tbody>
</table>

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s-IgA. No differences were observed between the contact lens-wearing groups and controls; patients with cystic fibrosis, however, had very high specific IgA responses (Tables 2, 3).

Specificity of the anti-\(P. \text{aeruginosa}\) antibodies was demonstrated by immunoblotting. A representative experiment indicates that tears of contact lens wearers react with various bands of an outer membrane protein preparation of the bacterium (Fig. 2). Negative controls included strips containing bacterial outer membranes in which tears were omitted in the whole procedure did not result in the appearance of detectable bands (data not shown).

To investigate whether the contact lens containers were contaminated with \(P. \text{aeruginosa}\), we investigated 56 storage cases using a special \(P. \text{aeruginosa}\) culture medium. None of them contained detectable \(P. \text{aeruginosa}\).

**DISCUSSION**

This study shows that a substantial number of persons lack detectable IgA antibodies against \(P. \text{aeruginosa}\) in their tears and may be susceptible to \(P. \text{aeruginosa}\) keratitis if they encounter this organism under circumstances in which the physiological condition of the cornea is compromised. Because we only used one isolate (O1 serotype) as a substrate for the detection of IgA antibodies against \(P. \text{aeruginosa}\), it may not be justified to extrapolate our findings to all strains of \(P. \text{aeruginosa}\). The O1 serotype, however, is one of the common strains involved in \(P. \text{aeruginosa}\) keratitis.\(^{18}\)

\(P. \text{aeruginosa}\) keratitis is the most frequent infectious complication during contact lens wear, and overnight wear appears to be the most important risk factor. Other factors involved in the pathogenesis of \(P. \text{aeruginosa}\) induced keratitis include direct in eye adhesion or contamination of the contact lens care system with the organism, a compromised corneal epithelium, and the absence of adequate specific and non-specific antibacterial factors in the tear film. Recent studies\(^{3,4}\) revealed that lens hygiene only has marginal effects on the development of microbial keratitis, which indicates that other factors make some persons wearing certain types of lenses uniquely susceptible to infection. During extended-wear contact lens use, the chance of compromising the cornea is greater\(^{23}\) than during daily wear. An interesting finding in the current study was that the anti-\(P. \text{aeruginosa}\) response was significantly lower in extended-wear users than it was in controls. This suggests that a compromised corneal physiology, in combination with a decreased local specific anti-bacterial immune response, may be the key factors in the pathogenesis of contact lens-associated keratitis.

The specificity of the local ocular IgA response has not received much attention until now, and most studies have dealt with the total IgA levels in tears during contact lens wear. S-IgA plays an important role in the prevention of bacterial adherence to mucosal tissues,\(^{41}\) and experimental studies have shown that stimulation of the local ocular immune response can ameliorate \(P. \text{aeruginosa}\) keratitis.\(^{25,26}\) Besides a specific immune response, ocular innate immune responses in the tear film—which include mucins, lysozyme, and lactoferrin—may play a role in the defense against microbial infection. In this study, we did not observe significant differences in total tear s-IgA between various groups of contact lens wearers and controls, which is in agreement with our previous studies using a high-performance liquid chromatography technique to quantitate tear IgA levels.\(^{27}\) However, other studies have reported markedly decreased\(^{28}\) and increased\(^{29}\) tear IgA levels in contact lens wearers. These discrepancies with our findings may have been caused either
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by differences in the sampling techniques as well as the method used to quantitate tear s-IgA. Sampling in our study was performed using a standardized technique and was carried out by the same trained observer throughout the study.

Analysis of IgA anti-P. aeruginosa antibodies has not yet been reported during contact lens wear. An earlier study addressed the ocular IgA response against P. aeruginosa in patients with cystic fibrosis and reported a much stronger specific IgA response in this patient group than in controls, a finding that could be confirmed by our study. The high anti-P. aeruginosa IgA response in patients with cystic fibrosis is attributable to the fact that the respiratory tract of these patients often is colonized by P. aeruginosa. In the current study, a huge range response was observed for the IgA anti-P. aeruginosa levels in the groups investigated, which may reflect the variability in the mucosal immune response among persons against this bacterium, a finding also reported earlier by others.

We could not detect Pseudomonas contamination in the contact lens carrying cases of patients visiting our contact lens clinics. This indicates that this population is not exposed to this antigen on a routine basis, and it may account for their low specific IgA response against P. aeruginosa.

The mechanisms responsible for the apparent lower anti-P. aeruginosa responses we observed in extended-wear contact lens users are not yet clear. One may consider the possibility that these lenses cover the entire cornea and part of the conjunctiva for prolonged periods of time. Mechanical or other barriers can prevent optimal antigen presentation to the dendritic cells of the peripheral cornea, which are thought to be involved in the initiation of the IgA response.

The lower anti-P. aeruginosa IgA response in the extended-wear contact lens users could have been caused by differences in total s-IgA in their tears. Although this group did show a trend toward lower s-IgA concentrations than the other groups, the differences were not statistically significant. We also could not detect differences in the tear production among the groups studied, which could account for possible s-IgA concentration differences. No correlation was observed between the amount of tears collected during the 1-minute collection period and the corresponding s-IgA concentration in the obtained sample.

The findings reported in this article suggest a role for a decreased specific IgA response in bacterial keratitis in extended-wear contact lens wearers. More vigorous testing of this hypothesis may be obtained by analyzing the local antibacterial response in a case-control study in patients with bacterial keratitis. If such testing confirms our hypothesis, it may well provide the rationale to implement preventive measures other than improving lens hygiene.

**Key Words**

contact lens, keratitis, *Pseudomonas aeruginosa*, secretory immunoglobulin A, tears

**Acknowledgment**

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**References**


