Supplemental Figures & Figure Legends

Figure S1. Flow cytometry analysis of HVEM expression in the HSV-1-infected murine cornea. (A) Representative histogram comparing HVEM expression in WT (red curve) and HVEM KO (black curve) corneas 3 dpi or (C) 14 dpi with HSV-1(17). Threshold for HVEM expression was set with an isotype control, FMO control, and HVEM KO sample from the same time point. (B) Representative dot plot of CD45 expression in live, HVEM⁺ cells from a WT sample 3 dpi or (D) 14 dpi.
Figure S2. Gating strategy for lymphoid cell populations. Whole corneas were assessed by flow cytometry. Cell populations were gated according to FSC-SSC, then restricted to singlets, live cells, CD45+ cells, CD3+ cells, and then were segregated into CD4 versus CD8 populations or NK1.1+ or NK1.1− populations. Cells negative for CD4, CD8, and NK1.1 were considered DN cells.
Figure S3. Gating strategy for myeloid cell populations. Whole corneas were assessed by flow cytometry. Cell populations were gated according to FSC-SSC, then restricted to singlets, live cells, and CD11b\(^+\) cells. Neutrophils were classified as the Ly6C\(^-\) Ly6\(^+\) population. The rest of cells were then separated into myeloid DCs, monocytes/macrophages, inflammatory monocytes/macrophages, and non-inflammatory monocytes/macrophages populations as shown above.
Figure S4. Treatment with IMPs significantly increases the number of PMN, tended to increase other leukocytic populations, in the spleens of C57BL/6 mice. WT mice were infected with 2.0x10^6 PFU/5 µl per eye after corneal scarification with HSV-1(17) and treated with either IMPs or vehicle (PBS) intravenously 3-7 dpi. Less than 24 hours after the last dose (day 8), the spleens of a subset of the group were harvested and prepared for flow. (A and B) Absolute number of the indicated cell types in WT treated or mock-treated mice (n = 4, means ± SEMs, two tailed t test with Holm-Sidak’s correction for multiple comparisons). *, P ≤ 0.05.